



Commodity risk assessment of *Fragaria* seeds

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Office for Risk assessment & Research (BuRO)

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Abstract

Following a request from the National Plant Protection Organisation of the Netherlands, the Office for Risk assessment & research (BuRO) conducted a Commodity risk assessment of *Fragaria* seeds for the European Union (EU). Plant pests (including pathogens) were identified that can be transmitted through seeds of *Fragaria*. For those seed-transmitted pests that are not regulated in the EU, it was evaluated whether they fulfil the criteria of a Union quarantine pest (EU-Q). Evidence of seed transmission in *Fragaria* was found for five viruses: Arabis mosaic virus (ArMV, *Nepovirus arabis*), beet ringspot virus (BRSV, *Nepovirus betae*), *Fragaria chiloensis* latent virus (FCiLV, *Iilarvirus FCILV*), raspberry ringspot virus (RpRSV, *Nepovirus rubri*), and strawberry necrotic shock virus (SNSV, *Iilarvirus SNSV*). Limited or inconclusive evidence for seed transmission was found for *Fragaria chiloensis* cryptic virus (FCiCV) and tomato ringspot virus (ToRSV, *Nepovirus lycopersici*). Three out of these seven viruses (BRSV, FCiLV and FCiCV) are not regulated in the EU. They do not qualify for a EU-Q status. BRSV may be quite common in the EU because of its long history of presence in Europe and broad host range. For FCiLV and FCiCV, no economic impact is expected if they were to establish in the EU. Three of the viruses (ArMV, RpRSV and ToRSV) are Union regulated non-quarantine pests, of which ArMV and RpRSV are regulated for plants for planting (including seeds) of *Fragaria*. SNSV is an EU-Q and special requirements are in place for plants for planting of *Fragaria*, but seeds are exempted from these requirements. No bacterial or fungal pathogens or plant-parasitic nematodes were identified for which seed transmission has been demonstrated in *Fragaria*. A few fungi that are known to infect the strawberry fruit have been found to be seed-borne in *Fragaria* seeds. Cultivated *Fragaria* plants have so far mainly been propagated vegetatively. Consequently, there is little experience available on transmission of pests through *Fragaria* seeds and more pests may be seed-transmitted in *Fragaria* than currently known. Seed transmission of plant-parasitic nematodes seems unlikely because they are not or rarely associated with fruit of *Fragaria*.

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1 Introduction

The National Plant Protection Organisation of the Netherlands has requested the Office for Risk assessment & research (BuRO) to conduct a commodity risk assessment of *Fragaria* (strawberry) seeds. This request was made because propagation by seed is becoming increasingly important in strawberry (Naktuinbouw, 2024; Bentvelsen, 2025). Therefore, it is important to know which pests (including pathogens) of *Fragaria* are seed-transmitted, so that measures can be established to prevent the introduction and spread of these pests through import and movement of *Fragaria* seeds in the European Union (EU).

The 'seeds' of *Fragaria* that are traded and sown are in fact so-called achenes. The edible fleshy strawberry fruit is an 'accessory fruit' with many tiny achenes (approximately 1 mm in length) on the outside (PlantAtlas, 2025). These achenes are dry fruits that do not split open to release the single seed (Ohio Plants, 2025). Pests that colonize the accessory fruit may, therefore, also be associated with the achenes. In this commodity risk assessment, the achenes will be referred to as 'seeds'.

The request included the following specific questions that are addressed in this commodity risk assessment:

- *which pests can be associated with Fragaria seeds (i.e. which pests are seed-borne in Fragaria)?*
- *which of those pests are seed-transmitted in Fragaria?*
- *what is the regulatory status in the EU of the pests that are seed-transmitted in Fragaria?*
- *do the seed-transmittable pests that are not regulated in the EU fulfil the criteria of a Union quarantine pest (EU-Q) (to be assessed by a pest categorization)?*

2 Data and methodologies

2.1 Terminology and abbreviations

2.1.1 Glossary of technical terms used in the document

Contaminating pest	A pest that is not seed-borne but can be present mixed with seeds in storage (FAO, 2021)
Pest	Any species, strain or biotype of plant, animal or pathogenic agent injurious to plants or plant products (FAO, 2021)
Plants for planting	Plants intended to remain planted, to be planted or to be replanted; 'plants' include seeds, in the botanical sense, other than those not intended for planting (Regulation (EU) 2016/2031) ¹
Seed-borne pest	A pest carried by seeds externally or internally that may or may not be transmitted to plants growing from these seeds and cause their infestation (FAO, 2021)
Seed-transmitted pest	A seed-borne pest that is transmitted via seeds directly to plants growing from these seeds and causes their infestation (FAO, 2021)

2.1.2 Abbreviations used in the document

EFSA	European Food Safety Authority https://www.efsa.europa.eu/
EPPO	European and Mediterranean Plant Protection Organisation

¹ Regulation (EU) 2016/2031 of the European Parliament of the Council of 26 October 2016 on protective measures against pests of plants. OJ L 317 23.11.2016, p. 4-104

EU-Q	Union quarantine pest (Regulation (EU) 2016/2031)
LPSN	List of Prokaryotic names with Standing in Nomenclature (Parte et al., 2020)
PZ-Q	Protected zone quarantine pest (Regulation (EU) 2016/2031)
RNQP	Union regulated non-quarantine pest (Regulation (EU) 2016/2031)

2.2 Identification of pests potentially associated with the commodity

A literature search was conducted to identify pests that have been reported to be associated with *Fragaria* seeds. Biological Abstracts 1969 to March 2025 and CAB abstracts 1973 to 2025 Week 14 were searched through Ovid® using the search string '(Fragaria OR strawberry) AND (seedborne OR seed-borne OR seed-transmitted OR (seed AND (transmission OR transmitted OR transmissible))' (in a preliminary search the term 'achene' did not yield more papers with relevant information). In addition, literature and databases were searched to identify regulated and non-regulated pests that infect *Fragaria* and may potentially be seed-borne (and seed-transmitted) in *Fragaria*. The following sources were used for specific groups of pests:

For regulated pests:

- EU-Qs, PZ-Qs and pests regulated by (national) temporary measures are listed in 'Register Q-organismen' on the website of the NVWA (NVWA, 2025). Pests that are known to infect plants of *Fragaria* were subsequently selected using the lists of host plants in the EPPO Global Database (EPPO, 2025b) and the CABI Compendium (CABI, 2025). RNQPs that are regulated for plants for planting of *Fragaria* were identified in Annex IV of Implementing regulation (EU) 2019/2072².

For all pest groups:

- EPPO Global Database (EPPO, 2025b): a search for all pests of '*Fragaria x ananassa*' and '*Fragaria*' listed in the database was made (a search for all pests of '*Fragaria*' only yielded a much shorter list of pests). The list of pests in the EPPO Global Database reflects the pest-specific information that has been produced or collected by EPPO, and is not complete and, therefore, additional searches were made per group of organisms (bacteria, fungi and oomycetes, nematodes and viruses and viroids) as described below.

For bacteria:

- Biological Abstracts 1969 to February 2025 and CAB abstracts 1973 to 2025 Week 11 were searched through Ovid® using the search string '(Fragaria OR strawberry) AND (Acidovorax OR Agrobacterium OR Liberibacter OR Clavibacter OR Curtobacterium OR Dickeya OR Erwinia OR Pantoea OR Paraburkholderia OR Pectobacterium OR Phlomobacter OR Pseudomonas OR Ralstonia OR Rhodococcus OR Spiroplasma OR Xanthomonas OR Xylophilus OR Xylella)'. This search string will identify all members of genera that contain bacteria which are currently EU-Qs or RNQPs in the EU or are mentioned in the EPPO A1/A2 lists.
- Biological Abstracts 1969 to February 2025 and CAB abstracts 1973 to 2025 Week 11 were searched through Ovid® using the search string '(Fragaria OR strawberry) AND (bacterial blight OR bacterial canker OR bacterial leaf blight OR bacterial leaf spot OR bacterial speck OR bacterial spot OR bacterial wilt)'. This search string focuses on typical symptoms related to bacterial infections.
- Only bacteria that were mentioned to be pathogenic in *Fragaria* in the identified papers were included in the list of pests.

For fungi and oomycetes:

- A list of fungal pathogens (including oomycetes) in a review of 'Fungal diseases of strawberry and their diagnosis' by Garrido et al. (2016). More fungi and oomycetes have been found associated with strawberry (USDA, 2025) but the list of Garrido et al. (2016) presumably

² Commission Implementing Regulation (EU) 2019/2072 of 28 November 2019 establishing uniform conditions for the implementation of Regulation (EU) 2016/2031 of the European Parliament and the Council, as regards protective measures against pests of plants. OJ L 319, 10.12.2019, p. 1-258

contains the most common fungal pathogens of strawberry and was considered sufficient for the objectives of the present study.

For nematodes (limited to foliar and stem nematodes):

- Biological Abstracts from 1969 to February 2025 and CAB abstracts from 1973 to 2025 Week 11 were searched through Ovid® for nematodes colonizing above ground plant parts of *Fragaria* other than *Ditylenchus dipsaci* using the search string '(Fragaria OR strawberry) AND (nematode OR nematodes) AND ((foliar OR stem) NOT dipsaci)'. *Ditylenchus dipsaci* was excluded from the search to keep the number of papers to a manageable number, but was included in the list of pests because it is known as a pest of *Fragaria* and is an RNQP for plants for planting of *Fragaria* (and *Ribes*).

For viruses and viroids:

- The inventory of viruses of *Cydonia* Mill., *Fragaria* L., *Malus* Mill., *Prunus* L., *Pyrus* L., *Ribes* L., *Rubus* L. and *Vitis* L. made by the EFSA Panel on Plant Health et al. (2019b). This study includes a list of all viruses and viroids of *Fragaria* that have been reported up to 19 March 2018.
- A literature search was conducted to identify new viruses and viroids reported to be associated with *Fragaria* since 19 March 2018. Therefore, Biological Abstracts was searched from January 2018 to March 2025 and CAB abstracts from January 2018 to 2025 Week 14 through Ovid® using the search string '(virus OR viroid) AND (Fragaria OR strawberry)'.

2.2.1 Organisms excluded

The following organisms were excluded from the assessment:

- Insects and mites: a preliminary literature search for seed-borne pests of *Fragaria* did not yield any insect or mite species. Furthermore, seeds of *Fragaria* are small; sizes in PlantAtlas (2025) are 1.3 mm in length and 1.1 mm in width. Therefore, any contaminating insects and mites are expected to be removed during harvesting and cleaning of the seeds.
- Fungi of the genera *Pichia*, *Saccharomyces* and *Zygosaccharomyces* that are known to cause postharvest rots in strawberry (Garrido et al., 2016). These pathogens (also known as yeasts) are unlikely to be seed-transmitted.
- Nematodes that only colonize or affect below ground parts and are, therefore, unlikely to be spread by seeds (e.g. *Meloidogyne*, *Pratylenchus* and *Xiphinema* spp.).
- Phytoplasmas: a few phytoplasmas are seed-transmitted according to Kirdat et al. (2022). These phytoplasmas are not known to infect plants of *Fragaria* (Calari et al., 2011; Kirdat et al., 2022). Seed-transmission of phytoplasmas has also been debated (Wei & Zhao, 2025). In addition, no publications of seed transmission were found for phytoplasmas in relation to strawberry in a pilot literature search (Biological Abstracts 1969 to March 2025 and CAB abstracts 1973 to 2025 Week 14 were searched for seed-transmitted pests of *Fragaria* using the search string '(Fragaria OR strawberry) AND (seedborne OR seed-borne OR seed-transmitted OR (seed AND (transmission OR transmitted OR transmissible))) in Ovid®).

2.2.2 Preferred names

The preferred name of each organism was verified using the EPPO Global Database (EPPO, 2025b). All identified nematodes were included in the EPPO Global Database but for fungi and oomycetes and bacteria that were not included in this database, MycoBank (2025) and the List of Prokaryotic names with Standing in Nomenclature (LPSN) (Parte et al., 2020) were used, respectively. For viruses and viroids the database of the International Committee on Taxonomy of Viruses (ICTV, 2025b) was used next to the EPPO Global Database. If the name of the virus or viroid was not included in these databases, the name mentioned in the publication was used.

2.2.3 Seed-borne and seed transmission

BuRO used the sources listed below to get information on whether a pest is seed-borne and seed-transmitted in *Fragaria* or other plants species. Pests, particularly bacteria, fungi and oomycetes, that are known to cause fruit diseases in *Fragaria* (other than *Pichia*, *Saccharomyces* and *Zygosaccharomyces* species; see section 2.2.1) were identified because these pests may be associated with the seeds.

In the assessment, a pest was considered seed-borne or seed-transmitted in *Fragaria* if the presence of the pest on/in seeds or seed transmission had been demonstrated in at least one *Fragaria* species. Cultivated *Fragaria* plants have so far mainly been propagated vegetatively and more pests may be seed-transmitted in *Fragaria* than currently known. Therefore, if no evidence was found for being seed-borne or seed-transmitted in *Fragaria*, information on other host plants of the pest was searched using the sources mentioned before for *Fragaria*. The search on other host plants was terminated when at least one other host plants was found in which seed transmission had been shown.

Sources:

- Datasheets of the pests in the EPPO Global Database and the CABI compendium and relevant references included in these datasheets (CABI, 2025; EPPO, 2025b).
- The following review papers and books:
 - 'New insights in the study of strawberry fungal pathogens' by Garrido et al. (2011) which included a list of fruit infecting fungi and oomycetes;
 - 'An annotated list of seed-borne diseases' by Richardson (1990);
 - 'Seed-borne plant virus diseases' by Sastry (2013);
 - 'Encyclopedia of plant viruses and viroids' by Sastry et al. (2019).
- Biological Abstracts 1969 to March 2025 and CAB abstracts 1973 to 2025 Week 14 were searched through Ovid® using the search strings:
 - '[name of the pest] and (seedborne or seed-borne or seed-transmitted or (seed and (transmission or transmitted or transmissible)))' to find any host of which the seeds have been found associated with the pest. For pests that yielded no results the search string '[name of the pest] and seed*' was used.
- Recent risk assessments of viruses infecting *Fragaria* including information on seed transmission (not available for other groups of pests):
 - pest categorisation of non-EU viruses of *Fragaria* L. from the EFSA Panel on Plant Health et al. (2019a),
 - scientific opinion on the risk to plant health posed by Arabis mosaic virus, raspberry ringspot virus, strawberry latent ringspot virus and tomato black ring virus to the EU territory with the identification and evaluation of risk reduction options (EFSA-Panel-on-Plant-Health, 2013).

Datasheets, review papers, reference books and risk assessments listed above were used to find original research papers that provide evidence for seed transmission of a particular pest.

3 Results and discussion

3.1 List of pests of *Fragaria*

All regulated and non-regulated organisms that were identified as pests of *Fragaria* and included for further study are listed in Annex A. Results are discussed below for each group of organisms:

- bacteria,
- fungi and oomycetes,
- nematodes,
- viruses and viroids.

3.2 Bacteria

No bacterial pathogens of *Fragaria* were identified that are known to be seed-borne in *Fragaria*.

Three bacterial pathogens of *Fragaria* were identified that are known to be seed-borne in other plant species: *Pantoea ananatis*, *Ralstonia pseudosolanacearum* and *Rhodococcus fascians*.

Pantoea ananatis is seed-borne in various host plants including onion (*Allium cepa*), maize (*Zea mays*) and Sudangrass (*Sorghum sudanense*) (Azad et al., 2000; Walcott et al., 2002; Rijavec et

al., 2007). Walcott et al. (2002) could recover *P. anatis* from onion seedlings. However, the seedlings were grown in Petri dishes and seed transmission studies under more natural conditions may be needed for confirmation that *P. anatis* is indeed seed-transmitted in onion. No further evidence was found for seed transmission of *P. anatis*.

Ralstonia pseudosolanacearum has more than 100 host plant species from different plant families and seed transmission has been reported in a few of its host plants (EFSA Panel on Plant Health et al., 2019). *Ralstonia pseudosolanacearum* (EU-Q) has been found to be seed-borne (and seed-transmitted) in groundnut (*Arachis hypogea*), tomato (*Solanum lycopersicum*) and eggplant (*S. melongena*) (Machmud & Middleton, 1990; Zhang et al., 1993; Sumithra et al., 2000; Dey et al., 2024). The bacterium has also been reported from *Capsicum* (pepper) seeds and seed transmission has been found through artificially infested *Capsicum* seeds (Preeti Aggarwal & Sood, 2005; Umesha et al., 2005). Seeds of *A. hypogea* appear to become infected by *R. pseudosolanacearum* (Machmud & Middleton, 1990; Zhang et al., 1993). The bacteria may, however, only be present externally on seeds of tomato and eggplant (and possibly pepper) because chemical seed treatments were able to eliminate the bacterium from tomato and eggplant seeds (Dey et al., 2024). It should be noted that there is some uncertainty whether the species reported in the publications referred to above is *R. pseudosolanacearum* or the related species *R. solanacearum*. They are both members of the *Ralstonia solanacearum* species complex that was split into different species including *R. solanacearum* and *R. pseudosolanacearum* in 2014 (Safni et al., 2014).

Rhodococcus fascians has many host plant species belonging to 87 genera and has been found to be seed-borne in some of these hosts (Putnam & Miller, 2007). It may also be seed-transmitted in these hosts. There is for example circumstantial evidence that *R. fascians* is seed-transmitted in *Lathyrus odoratus*. In preliminary experiments on the infectiousness of fasciation symptoms, Tilford (1936) found that a particular seed lot of *L. odoratus* always produced some diseased plants; the use of disinfested seeds from this lot resulted in no diseased plants. Lacey (1939) found two out of 54 seedlings of *Schizanthus retusus* infected by *R. fascians* after sowing seeds in sterilized soil.

One bacterial pathogen was identified that causes a fruit disease in *Fragaria*: *Erwinia pyrifoliae*. This pathogen may also be present on seeds from infected plants because the seeds (achenes) are present on the accessory fruit (see the 'Introduction'). *Erwinia pyrifoliae* can be transported by honey bees (Wenneker, 2022) but seed transmission has not been reported.

3.3 Fungi and oomycetes

3.3.1 List of fungi and oomycetes

A list of fungi and oomycetes that infect *Fragaria* is provided by Garrido et al. (2016). Some of these fungi and oomycetes were, however, excluded from the assessment because of uncertainty about the host status of *Fragaria*:

- *Peronospora potentillae*: Garrido et al. (2016) refer to Choi et al. (2009) for *P. potentillae* being a pathogen of *Fragaria*. However, Choi et al. (2009) describe other *Peronospora* species which infect *Salvia* and no evidence was provided that *P. potentillia* infects *Fragaria*. Douglas Gubler & Converse (1993) indicate *P. fragariae* as a synonym of *P. potentillia*, but *P. sparsa* is currently the preferred name of *P. fragariae* according to EPPO (2025b) and *Fragaria* is not included in the host list of *P. sparsa* in CABI (2020) (EPPO (2025b) does not provide a host list of *P. sparsa*).
- *Sphaeropsis malorum*: *S. malorum* is known as a pathogen of apple (*Malus domestica*) (Grove et al., 2003). No studies were found on *S. malorum* affecting *Fragaria*. Douglas Gubler & Converse (1993) indicate *Botryosphaeria obtusa* (current preferred name: *Diplodia serrata*) as a teleomorph of *S. malorum*. However, *Fragaria* is not listed as a host of *Diplodia serrata* either (Reeder, 2020).

Sphaerotheca macularis was replaced by *Podosphaera aphanis*:

- Garrido et al. (2016) list *Sphaerotheca macularis* as the causal agent of powdery mildew of *Fragaria*. However, the current preferred name of *S. macularis* is *Podosphaera macularis* which is not known as a pest of *Fragaria* (CABI, 2021d). *Podosphaera aphanis* is currently known as the causal agent of powdery mildew in *Fragaria* (CABI, 2021e) and was included in the list of species (Annex A). This species is an RNQP for plants for planting of *Fragaria* (Implementing Regulation (EU) 2019/2072).

Some *Colletotrichum* species were added to the list of Garrido et al. (2016) due to changes in the systematics of this genus:

- Three *Colletotrichum* species, *C. acutatum*, *C. gloeosporioides* and *C. fragariae*, are listed by Garrido et al. (2016). *Colletotrichum acutatum* and *C. gloeosporioides* are, however, species complexes (Rose & Damm, 2024). The species complex *Colletotrichum acutatum* s.l. (= *sensu lato*) is an RNQP for plants for planting of *Fragaria* (the EPPO-code 'COLLAC' in the EU-legislation refers to the species complex). For these reasons, BuRO included the following three categories within the genus *Colletotrichum*: *C. fragariae*, *Colletotrichum acutatum* s.l. and *Colletotrichum* spp. other than *C. acutatum* s.l. and *C. fragariae*.

3.3.2 Seed-borne and seed transmission

No plant pathogenic fungi or oomycetes were identified as being seed-transmitted in *Fragaria* but a few plant pathogenic fungi and oomycetes have been reported from *Fragaria* seeds (i.e. being seed-borne in *Fragaria*). CABI (2021f) states that *Colletotrichum* spp. are frequently isolated from *Fragaria* seeds and Naktuinbouw (pers. comm., 5 November 2025) has detected *Colletotrichum acutatum* s.l., *Colletotrichum gloeosporioides* s.l., *Neopestalotiopsis* sp., en *Phytophthora cactorum* on/in *Fragaria* seeds by TaqMan real-time PCR. These pathogens are all known to infect the fruit (and other plant parts) of *Fragaria* and more fungi and oomycetes that infect the fruit may be expected on the seeds. The majority of fruit-infecting species listed by Garrido et al. (2011) are also widespread in the EU and other parts of the world (Table 1). From the (groups of) fungi and oomycetes listed in Table 1, *C. fragariae* is the only species that is not known to be present in the EU. Many species that can infect *Fragaria* including species that affect the strawberry fruit are known to be seed-borne (and seed-transmitted) in other plant species (Table 2).

There are more fungi and oomycetes that can affect *Fragaria* fruit than those listed by Garrido et al. (2011), e.g. *Pilidium lythri* (previously named *P. concavum*) and *Neurospora dictyophora* (Lopes et al., 2010; Debode et al., 2011; Rivera et al., 2024). Under specific conditions, especially the fleshy fruit of *Fragaria* may be susceptible to various kinds of fungi, including species that are generally not known as plant pathogens. For example, Rivera et al. (2024) attributed fruit infections of *N. dictyophora* to poor ventilation and high temperatures in greenhouses in Mexico. Such organisms may also be associated with the seeds, but seed transmission seems unlikely for organisms that are only known to colonize the fruit and no other plant tissues. Fungi and oomycetes that are present on the fruit may be removed by disinfection of the seeds prior to sowing (Miller et al., 1992; El Hamdouni et al., 2001; Galvão et al., 2014).

Table 1. Fungi and oomycetes that cause fruit diseases in *Fragaria*: pest status in the EU according to (CABI, 2025) unless stated otherwise.

Species	Pest status in the EU (and other parts of the world)
<i>Alternaria tenuissima</i>	Reported in 5 EU member states; cosmopolitan (CABI, 2021h)
<i>Aspergillus niger</i>	Widespread in the EU (≥ 10 EU member states), cosmopolitan,
<i>Athelia rolfsii</i>	Widespread in the EU, cosmopolitan
<i>Botrytis cinerea</i>	Widespread in the EU, cosmopolitan
<i>Colletotrichum acutatum sensu lato</i>	Widespread in the EU, cosmopolitan
<i>Colletotrichum fragariae</i>	Not reported in the EU
<i>Colletotrichum</i> spp. other than <i>C. acutatum sensu lato</i> and <i>C. fragariae</i>	Cosmopolitan (Rose & Damm, 2024)
<i>Coniella straminea</i>	Cosmopolitan (Alvarez et al., 2016)
<i>Gnomoniopsis comari</i>	EU: Finland, Germany (MycoBank, 2025) Outside the EU: Switzerland (MycoBank, 2025)
<i>Mucor</i> spp.	<i>Mucor</i> spp. in general cause fruit rot, are cosmopolitan and are widespread in the EU
<i>Penicillium</i> spp.	<i>Penicillium</i> spp. in general cause fruit rot, are cosmopolitan and are widespread in the EU
<i>Pestalotiopsis longisetula</i>	Cosmopolitan (Van Hemelrijck et al., 2016; EPPO, 2021)

Species	Pest status in the EU (and other parts of the world)
<i>Phytophthora cactorum</i>	Widespread in the EU, cosmopolitan
<i>Phytophthora citrophthora</i>	Widespread in the EU, cosmopolitan
<i>Phytophthora nicotianae</i> var. <i>nicotianae</i>	Cosmopolitan (Waterhouse & Waterston, 1964)
<i>Ramularia grevilleana</i>	EU: Ireland, Poland (CABI, 2019), Germany (Richter & Jage, 2003) Outside the EU: India, Japan, United States (CABI, 2019), Iran (Heydari et al., 2017)
<i>Rhizoctonia fragariae</i>	EU: Italy, Spain Outside the EU: Egypt and the United States
<i>Rhizoctonia solani</i>	Cosmopolitan (Akber et al., 2023)
<i>Rhizopus sexualis</i>	Reported from Japan and United Kingdom (Lunn, 1977) but may have a wider distribution because <i>Rhizopus</i> species are generally considered cosmopolitan
<i>Rhizopus stolonifer</i>	Cosmopolitan
<i>Sclerotinia sclerotiorum</i>	Widespread in the EU, cosmopolitan
<i>Stagonospora fragariae</i>	Little information found: records in Europe in France (Fungorum, 2025) and Scotland (United Kingdom) (NBNAtlas, 2025)

Table 2. Seed transmission (ST) of plant pathogenic fungi and oomycetes of *Fragaria* found to be seedborne in other plant species (T = shown to be transmitted through seeds including indirect transmission via the environment, TI = transmission found after inoculation of seeds (transmission uncertain under natural conditions), NE = no evidence found for seed transmission).

Fungus/oomycete	Plant species¹	ST	References
<i>Alternaria alternata</i>	Various plant species	T	(Kgatle et al., 2018; CABI, 2021a)
<i>Alternaria tenuissima</i>	<i>Triticum aestivum</i> (wheat)	TI	(Perello & Larran, 2013)
<i>Aspergillus niger</i>	<i>Allium cepa</i> (onion)	T	(Hayden & Maude, 1992)
<i>Athelia rolfsii</i>	Various plant species	NE	(CABI, 2021b)
<i>Boeremia lycopersici</i>	<i>Solanum lycopersicum</i> (tomato)	T	(Phillips, 1956; Derbyshire, 1960)
<i>Botrytis cinerea</i>	<i>Cicer arietinum</i> (chickpea)	T	(Burgess et al., 1997)
<i>Cladosporium</i> spp.	<i>Phaseolus vulgaris</i> (bean), <i>Spinacia oleracea</i> (spinach)	T	(Hernandez-Perez, 2005; Guimarães et al., 2014)
<i>Colletotrichum</i> spp.	<i>Capsicum annuum</i> (pepper), <i>Phaseolus vulgaris</i> (bean)	T	(Tu, 1983; Lee, 1995; El-Ammari et al., 1997)
<i>Fusarium oxysporum</i>	Various plant species	T	(Bassi & Goode, 1978; Collins, 2016; Petkar & Ji, 2017)
<i>Fusarium sambucinum</i>	<i>Allium cepa</i> (onion)	T	(Koycu & Ozer, 1997)
<i>Macrophomina phaseolina</i>	<i>Phaseolus vulgaris</i> (common bean)	T	(Parsa et al., 2016)

Fungus/oomycete	Plant species¹	ST	References
<i>Mucor</i> spp.	<i>Vigna unguiculata</i> (cowpea)	NE	(Pachoute et al., 2024)
<i>Paraconiothyrium fuckelii</i>	<i>Capsicum frutescens</i> (chili pepper)	NE	(Chigoziri & Ekefan, 2013)
<i>Penicillium</i> spp.	<i>Zea mays</i> (maize)	T	(CABI, 2021i)
<i>Phytophthora cactorum</i>	<i>Carthamus tinctorius</i>	NE	(Zad, 1992)
<i>Phytophthora citrophthora</i>	<i>Citrus</i>	T	(CABI, 2021g)
<i>Phytophthora cryptogea</i>	<i>Cichorium</i> (chicory), <i>Spinacia oleracea</i> (spinach)	NE	(Gullino et al., 2012) ²
<i>Phytophthora megasperma</i>	<i>Pisum sativum</i> (pea)	NE	(Ozgonen & Gulcu, 2011)
<i>Phytophthora nicotianae</i> var. <i>nicotianae</i>	<i>Capsicum annuum</i> (sweet pepper)	NE	(Bhardwaj et al., 1987)
<i>Pythium ultimum</i>	<i>Pisum sativum</i> (pea)	NE	(El-Nagerabi et al., 2000)
<i>Rhizoctonia solani</i>	<i>Gossypium hirsutum</i> (Cotton)	TI	(Khatun et al., 2023)
<i>Rhizopus stolonifer</i>	<i>Zea mays</i> (maize)	T	(dos Santos e Silva et al., 2019; CABI, 2021c)
<i>Sclerotinia sclerotiorum</i>	Various species	T ³	(Saharan & Mehta, 2008)
<i>Verticillium alboatrum</i> sensu lato	<i>Medicago sativa</i> (alfalfa)	T	(Christen, 1982; Gilbert & Peaden, 1988)
<i>Verticillium dahliae</i>	<i>Helianthus annuus</i> (sunflower)	T	(Sackston & Martens, 1959)

¹ The species may be seed-borne (and seed-transmitted) in additional host plants, but no full literature search was performed to identify all host plants.

² No details were provided how *P. cryptogea* was confirmed to be present on the seeds of chicory and spinach.

³ Generally considered to be seed-transmitted in various crops but no experimental evidence found through naturally infected seeds.

3.4 Nematodes

Plant-parasitic nematodes that affect above-ground plant parts of *Fragaria* (*Ditylenchus dipsaci* and various *Aphelenchoides* spp.) are seed-borne and seed-transmitted in various plant species, but they are not known to be seed-borne in *Fragaria* (Table 3). In addition to the *Aphelenchoides* species listed in Table 3, *A. smolae* has been found associated with *Fragaria* (Subbotin, 2024). However, it is not known whether this species actually causes any disease in *Fragaria* or other plant species.

According to Pscheidt & Ocamb (2025), *D. dipsaci* is rarely found in strawberry fruit. In addition, the seeds of *Fragaria* may be too small to carry *Ditylenchus dipsaci*. The seeds of *Fragaria* that are measure approximately 1.3 mm (examples of sizes in PlantAtlas (2025) are: *Fragaria x ananassa* length 1.3 and width 1.1 mm, *F. moschata* length 1.3 and width 1 mm, *F. vesca* length 1.3 and width 0.9 mm), while J4-juveniles of *D. dipsaci* (the survival stage in dry seeds) are approximately 1.0 mm long (Esquibet et al., 1998). For these reasons, seed transmission of *D. dipsaci* in *Fragaria* seems unlikely.

Aphelenchoides species are generally smaller than *D. dipsaci* which makes their presence in small seeds more likely. However, *Aphelenchoides* species are known to feed on leaves and buds of *Fragaria*, but fruit of *Fragaria* is not known to carry the nematodes (FAO, 2016; CABI, 2022; Bozbuga et al., 2023). Therefore, association of *Aphelenchoides* spp. with *Fragaria* seeds seems unlikely.

Table 3. Plant-parasitic nematodes of *Fragaria* known to be seed-transmitted in plant species other than *Fragaria*

Nematode	Host plants ¹	Reference
<i>Aphelenchoides besseyi</i>	<i>Oryza sativa</i> (rice)	(EPPO, 2015)
<i>Aphelenchoides blastophthorus</i>	<i>Callistephus chinensis</i>	(Burckhardt, 1972)
<i>Aphelenchoides fragariae</i> ²	Forage grasses	(Favoreto et al., 2011)
<i>Aphelenchoides ritzemabosi</i>	<i>Aster</i> (aster), <i>Callistephus chinensis</i> , <i>Salvia splendens</i> (scarlet sage)	(Burckhardt, 1972; Chalanska et al., 2011; Blancard, 2014)
<i>Ditylenchus dipsaci</i>	<i>Allium</i> spp., <i>Beta vulgaris</i> (beet), <i>Cucumis melo</i> (melon), <i>Dipsacus</i> spp. (teasel), <i>Medicago sativa</i> (lucerne/alfalfa), <i>Pisum sativum</i> (pea), <i>Vicia</i> spp.	(Caubel, 1983; Hajihassani et al., 2016; Kumar & Gupta, 2020)

¹ The listed host plants are examples and no full literature search was performed to identify all host plants in which the pest is known to be seed-borne.

² No experimental evidence found, but because of biological similarities with the other *Aphelenchoides* spp. considered to be transmitted by seeds.

3.5 Viruses and viroids

Fourty-three viruses and viroids were identified that have been reported to infect *Fragaria* (Annex A). Two of the viruses are now considered phantom agents. For three viruses, confirmation is needed that *Fragaria* is a true (natural) host plant. This includes raspberry bushy dwarf virus (RBDV, *Idaeovirus rubi*), for which seed transmission has been shown after experimental inoculation of plants (Annex B). However, because there is no conclusive evidence that *Fragaria* is a natural host, RBDV was not included in the list of viruses with evidence for seed transmission in *Fragaria*. *Fragaria* was, however, considered a natural host of the remaining 38 viruses and viroids. For five out of these 38 viruses and viroids (Arabis mosaic virus, beet ringspot virus, *Fragaria chiloensis* latent virus, raspberry ringspot virus, strawberry necrotic shock virus), evidence was found that they are seed-transmitted in *Fragaria* (Table 4, Annex B). For two other viruses (*Fragaria chiloensis* cryptic virus, tomato ringspot virus), the current evidence was considered limited or inconclusive (Table 5, Annex B). The EU regulatory status of these seven viruses is discussed in section 3.5.1. This section also discusses the need of a pest categorisation of the non-regulated viruses. Section 3.5.2 lists the viruses and viroids for which no evidence was found for seed transmission in *Fragaria* but for which evidence was found in other plant species.

3.5.1 Regulatory status of viruses with (inconclusive or limited) evidence for seed transmission

Arabis mosaic virus and raspberry ringspot virus are RNQPs in the EU for plants for planting of *Fragaria* (including seeds) (Table 4).

Beet ringspot virus (BRSV), *Fragaria chiloensis* latent virus (FCiLV) and *Fragaria chiloensis* cryptic virus (FCiCV) are not regulated in the EU (Tables 4,5):

- BRSV has a broad host range including potato, sugar beet, turnip, many ornamental species and weeds (Harrison, 1957; Kis et al., 2017; Jordan et al., 2019; Motsar et al., 2025). In the Netherlands, BRSV has been detected in plants of *Cucurbita pepo* (pers. comm. NWWA-NIVIP, October 2025). BRSV was formerly considered a strain of tomato black ring virus (TBRV), a virus that is an RNQP for plants for planting other than seeds of *Fragaria*, *Prunus avium*, *Prunus cerasus* and *Rubus* (Implementing Regulation (EU) 2019/2072, Annex IV, Part J). BRSV has been present in Europe for many decades but there is little information about its distribution in the EU (Fowkes et al., 2022). In the EU, BRSV has been isolated from plants in at least three EU member states (Germany, Hungary and the Netherlands) (Fowkes et al., 2022; Jones et al., 2022; Motsar et al., 2025). Presumably, it has a wider distribution because of its long history of presence in Europe and its broad host range. In the past, isolates that have been identified as TBRV may actually have been BRSV. In view of the above, BuRO does not consider this virus to fulfil the criteria of a Union quarantine pest and no pest categorisation was made³.
- FCiLV and FCiCV: EFSA Panel on Plant Health et al. (2019a) conducted a pest categorisation for FCiLV and FCiCV and concluded that these viruses do not qualify for a Union quarantine status because no economic impact was expected if the viruses were to become established in the EU. BuRO agrees with this conclusion.

Tomato ringspot virus (ToRSV) is an RNQP in the EU for plants for planting other than pollen and seeds of *Malus*, *Prunus* and *Vaccinium* and plants for planting other than pollen of *Rubus* but is not regulated for *Fragaria* (Implementing Regulation (EU) 2019/2072, Annex IV, Part J; Table 5).

Strawberry necrotic shock virus, is an EU-Q and is (by definition) regulated for all plants for planting (Table 4). Special requirements are in place for plants for planting of *Cydonia*, *Fragaria*, *Malus*, *Prunus*, *Pyrus*, *Ribes* and *Rubus* to reduce the risk of SNSV but seeds are exempted from these requirements (Annex VII of Implementing Regulation (EU) 2019/2072, point 45).

Table 4. Regulatory status of viruses with evidence for seed transmission in *Fragaria* (see Annex B for details and references)

Virus	Binomial name	Regulatory status	Regulated for plants for planting other than seeds of <i>Fragaria</i>?	Regulated for seeds of <i>Fragaria</i>?
Arabis mosaic virus	<i>Nepovirus arabis</i>	RNQP	Yes	Yes
Beet ringspot virus	<i>Nepovirus betae</i>	Not regulated	No	No
Fragaria chiloensis latent virus	<i>Iilarvirus FCILV</i>	Not regulated	No	No
Raspberry ringspot virus	<i>Nepovirus rubri</i>	RNQP	Yes	Yes
Strawberry necrotic shock virus	<i>Iilarvirus SNSV</i>	EU-Q	Yes	Yes (but seeds are exempted from the special requirements) ¹

¹ Special requirements in Annex VII of Implementing Regulation (EU) 2019/2072, point 45

³ One of the criteria of an EU-Q is that the pest is not present in the EU or, if present, is not widely distributed (Articles 3-4 in Regulation (EU) 2016/2031 of 23.11.2016, OJ 317, 4-104).

Table 5. Regulatory status of viruses with limited or inconclusive evidence for seed transmission in *Fragaria* (see Annex B for details and references)

Virus	Binomial name	Regulatory status	Regulated for plants for planting of <i>Fragaria</i>?	Regulated for seeds of <i>Fragaria</i>?
Fragaria chiloensis cryptic virus	-	Not regulated	No	No
Tomato ringspot virus	<i>Nepovirus lycopersici</i>	RNQP	No	No

3.5.2 Viruses with evidence for seed transmission in other plant species

Three viruses (apple mosaic virus, tobacco streak virus and tomato black ring virus) have been listed as being seed-transmitted in a review of strawberry viruses (Tzanetakis & Martin, 2013). However, no supporting evidence of seed transmission in *Fragaria* was found for these viruses. For these three viruses, another four viruses and one viroid, evidence for seed transmission was found in other host plants (Table 6, Annex B).

Table 6. Viruses with no evidence of seed transmission in *Fragaria* but evidence of seed transmission in other host plants (see Annex B for details and references)

Virus/viroid	Binomial name	Host plants with evidence of seed transmission¹
Apple mosaic virus	<i>Iilarvirus ApMV</i>	<i>Corylus avellana</i> (hazelnut)
Cucumber mosaic virus	<i>Cucumovirus CMV</i>	<i>Capsicum annuum</i> (pepper), <i>Spinacia oleraceae</i> (spinach), <i>Vigna unguiculata</i> (cowpea)
Hop stunt viroid	<i>Hostuviroid impedihumuli</i>	<i>Vitis vinifera</i> (grape), <i>Prunus domestica</i> (plum)
Olive latent virus 1	<i>Alphanecrovirus oleae</i>	<i>Olea europaea</i> (olive)
Spinach latent virus	<i>Iilarvirus SLV</i>	<i>Chenopodium quinoa</i> (quinoa), <i>Nicotiana clevelandii</i> (Cleveland's tobacco) and <i>Spinacia oleracea</i> (spinach)
Strawberry latent ringspot virus	<i>Stralarivirus fragariae</i>	<i>Chenopodium quinoa</i> (quinoa), <i>Petroselinum crispum</i> (parsley), <i>Stellaria media</i> (common chickweed)
Tobacco streak virus	<i>Iilarvirus TSV</i>	<i>Glycine max</i> (soja), <i>Phaseolus vulgaris</i> (common bean), <i>Verbesina encelioides</i> (American dogweed), <i>Vigna angularis</i> (red mung bean)
Tomato black ring virus	<i>Nepovirus nigranuli</i>	<i>Lactuca sativa</i> (lettuce), <i>Sambucus nigra</i> (elder)

¹ The virus or viroid may be seed-transmitted in more host plants, but no full literature search was done to identify all host plants in which the pest has been found to be seed-transmitted.

4 Discussion and conclusions

Evidence of seed transmission in *Fragaria* was found for five viruses:

- Arabis mosaic virus (ArMV, *Nepovirus arabis*),
- beet ringspot virus (BRSV, *Nepovirus betae*),
- *Fragaria chiloensis* latent virus (FCiLV, *Iilarvirus FCiLV*),
- raspberry ringspot virus (RRV, *Nepovirus rubri*),
- strawberry necrotic shock virus (SNSV, *Iilarvirus SNSV*).

In addition, limited or inconclusive evidence of seed transmission in *Fragaria* was found for two other viruses:

- *Fragaria chiloensis* cryptic virus (FCiCV)
- tomato ringspot virus (ToRSV, *Nepovirus lycopersici*)

ArMV and RRV are RNQPs. Both viruses are regulated for seeds of *Fragaria* (and other plants for planting).

BRSV, FCiLV and FCiCV are not regulated in the EU. BRSV does not qualify for a Union quarantine status because it is already present in the EU where it is presumably quite common (i.e. does not have a limited distribution). BRSV may qualify for an RNQP status, but testing against RNQP-criteria was not within the scope of the present assessment. FCiLV and FCiCV do not qualify for a Union quarantine status because no economic impact is expected if these viruses would become established in the EU.

SNSV has a Union quarantine status and special requirements are in place for plants for planting of *Fragaria*. Seeds are currently exempted from these requirements. ToRSV is an RNQP but not for plants for planting (including seeds) of *Fragaria*.

For seven additional viruses that are known to naturally infect *Fragaria*, evidence was found for seed transmission in other hosts than *Fragaria*, while evidence is lacking for seed transmission in *Fragaria*. Three of these viruses (apple mosaic virus, tobacco streak virus and tomato black ring virus) have been listed as being seed-transmitted in *Fragaria* by Tzanetakis & Martin (2013) but without reference to original experimental evidence. It should, however, be taken into account that more viruses may be seed-transmitted in *Fragaria* than those for which experimental evidence was found in the present assessment. This may especially be the case for viruses that belong to the genera *Nepovirus* and *Iilarvirus* including apple mosaic virus, tobacco streak virus and tomato black ring virus. Seed transmission of nepoviruses is very common in herbaceous hosts (ICTV, 2026c) and many carlaviruses are also known to be seed-transmitted (Tables 4,6).

No evidence of seed transmission in *Fragaria* was found for plant-parasitic nematodes. Several plant-parasitic nematodes of *Fragaria* were identified as being seed-transmitted in other plant species. However, seed transmission of these nematodes in *Fragaria* seems unlikely because they are rarely present on the fruit and for some nematodes the seeds may be too small.

Evidence of seed transmission in *Fragaria* was neither found for bacterial or fungal pathogens. A few fungal species that colonize fruits of *Fragaria* have been reported from *Fragaria* seeds, but are only known as seed-borne. Many other fungi and oomycetes and the bacterium *E. pyrifoliae* can also colonize the fruits of *Fragaria*. These pathogens may, therefore, also be seed-borne.

Propagation of strawberries by seed is becoming increasingly important. So far, cultivated *Fragaria* plants have mainly been propagated vegetatively. Consequently, there is limited experience with transmission of pathogens through *Fragaria* seeds. Additional pathogens may thus be identified as seed-transmitted in *Fragaria* over time, partly because several of the pathogens of *Fragaria* have been identified as seed-transmitted in other plant species.

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5 References

- Abdullahi I, Ikotun T, Winter S, Thottappilly G & Atiri G, 2001. Investigation on seed transmission of cucumber mosaic virus in cowpea. *African Crop Science Journal*, 9 (4), 677-684.
- Akber MA, Mubeen M, Sohail MA, Khan SW, Solanki MK, Khalid R, Abbas A, Divvela PK & Zhou L, 2023. Global distribution, traditional and modern detection, diagnostic, and management approaches of *Rhizoctonia solani* associated with legume crops. *Frontiers in Microbiology*, 13, 1091288.
- Ali A & Kobayashi M, 2010. Seed transmission of Cucumber mosaic virus in pepper. *Journal of virological methods*, 163 (2), 234-237.
- Allen WR, Davidson T & Briscoe M, 1970. Properties of a strain of Strawberry latent ringspot virus isolated from sweet cherry growing in Ontario. *Phytopathology*, 60 (8), 1262-1265.
- Alvarez L, Groenewald J & Crous P, 2016. Revising the Schizoparmaceae: *Coniella* and its synonyms *Pilidiella* and *Schizoparme*. *Studies in Mycology*, 85 (1), 1-34.
- Azad HR, Holmes GJ & Cooksey DA, 2000. A new leaf blotch disease of sudangrass caused by *Pantoea ananas* and *Pantoea stewartii*. *Plant Disease*, 84 (9), 973-979. Available online: <https://doi.org/10.1094/PDIS.2000.84.9.973>
- Bassi A, Jr & Goode M, 1978. *Fusarium oxysporum* f. sp. *spinaciae* seedborne in spinach. *Plant Disease Reporter*, 62, 203-205.
- Bellardi M & Bertaccini A, 1992. Strawberry latent ringspot virus (SLRV) infection and parsley seed germination.
- Bentvelsen G, 2025. Aardbei F1-hybriden uit zaad. *Gewasbescherming*, Jaargang 56 (4), 128-129.
- Bhardwaj SS, Sharma SL & Tyagi SNS, 1987. Perpetuation of *Phytophthora nicotianae* var. *nicotianae* in bell pepper seeds. *Indian Journal of Plant Pathology*, 5 (1).
- Blancard D, 2014. *Aphelenchoides ritzemabosi*, biology and epidemiology [Web page]. INRA. Available online: <https://ephytia.inra.fr/en/C/10939/Tobacco-Biology-and-Epidemiology#:~:text=Aphelenchoides%20ritzemabosi%20can%20also%20be%20spread%20by%20seed%20in%20a%20sterile%20medium,&text=The%20nematode%20and%20the%20disease,is%20favoured%20by%20rainy%20periods>. [Accessed: 10-04-2025].
- Bos L, Huttinga H & Maat D, 1979. Parsley latent virus, a new and prevalent seed-transmitted, but possibly harmless virus of *Petroselinum crispum*. *Netherlands Journal of Plant Pathology*, 85 (3), 125-136.
- Bos L, Huttinga H & Maat D, 1980. Spinach latent virus, a new ilarvirus seed-borne in *Spinacia oleracea*. *Netherlands Journal of Plant Pathology*, 86 (2), 79-98.
- Bozbuga R, Uluisik S, Kara PA, Yuceer S, Gunacti H, Guler PG, Ince E, Yildiz HN & Tetik O, 2023. Pests, Diseases, Nematodes, and Weeds Management on Strawberries. In, *Recent Studies on Strawberries*. IntechOpen.
- Bradbury J, 1967. *Corynebacterium fascians*. *Descriptions of Fungi and Bacteria*, (13), Sheet 121.
- Bradbury J, 1991. *Xylella fastidiosa*. *Descriptions of Fungi and Bacteria*, (105), Sheet 1049.
- Braun A & Keplinger J, 1973. Seed transmission of tomato ringspot virus in raspberry. *Plant Disease Reporter*, 57 (5), 431-432.
- Burckhardt F, 1972. New observations on the presence of *Aphelenchoides* spp. in seed of *Callistephus chinensis*. *Nachrichtenblatt des Deutschen Pflanzenschutzdienstes Braunschweig*, 24 (9), 132-133.
- Burgess DR, Bretag T & Keane PJ, 1997. Seed-to-seedling transmission of *Botrytis cinerea* in chickpea and disinfestation of seed with moist heat. *Australian Journal of Experimental Agriculture*, 37 (2), 223-229.
- CABI, 2019. *Mycosphaerella fragariae* (leaf spot of strawberry) [Web page]. Available online: <https://www.cabidigitallibrary.org/doi/full/10.1079/cabicompendium.35280>
- CABI, 2020. *Peronospora sparsa* (downy mildew) [Web page]. Available online: <https://www.cabidigitallibrary.org/doi/full/10.1079/cabicompendium.39730#sec-14> [Accessed: 23-05-2025].
- CABI, 2021a. *Alternaria alternata* (alternaria leaf spot) [Web page]. CAB International. Available online: <https://www.cabidigitallibrary.org/doi/full/10.1079/cabicompendium.4480#sec-9>
- CABI, 2021b. *Athelia rolfsii* (Sclerotium rot) [Web page]. CAB International. Available online: <https://www.cabidigitallibrary.org/doi/full/10.1079/cabicompendium.49155#sec-18>
- CABI, 2021c. *Rhizopus stolonifer* (bulb rot) [Web page]. Available online: <https://www.cabidigitallibrary.org/doi/full/10.1079/cabicompendium.47665#sec-10>
- CABI, 2021d. *Podosphaera macularis* (powdery mildew of hop) [Web page]. Available online: <https://www.cabidigitallibrary.org/doi/full/10.1079/cabicompendium.50923#sec-7>
- CABI, 2021e. *Podosphaera aphanis* (powdery mildew of strawberry) [Web page]. Available online: <https://www.cabidigitallibrary.org/doi/full/10.1079/cabicompendium.42645>

- CABI, 2021f. *Colletotrichum fragariae* (anthracnose of strawberry) [Web page]. Available online: <https://www.cabidigitallibrary.org/doi/10.1079/cabicompendium.14907> [Accessed: 23-05-2025].
- CABI, 2021g. *Phytophthora citrophthora* (brown rot of citrus fruit) [Web page]. Available online: <https://www.cabidigitallibrary.org/doi/full/10.1079/cabicompendium.40958#sec-16> [Accessed: 23-05-2025].
- CABI, 2021h. *Alternaria tenuissima* (nailhead spot of tomato) [Web page]. Available online: <https://doi.org/10.1079/cabicompendium.4516>
- CABI, 2021i. *Penicillium* (penicillium ear rot) [Web page]. Available online: <https://www.cabidigitallibrary.org/doi/full/10.1079/cabicompendium.39561>
- CABI, 2022. *Aphelenchoides fragariae* (strawberry crimp nematode) [Web page]. Available online: <https://www.cabidigitallibrary.org/doi/full/10.1079/cabicompendium.6381#sec-10>
- CABI, 2024. Tobacco ringspot virus [Web page]. Available online: <https://www.cabidigitallibrary.org/doi/full/10.1079/cabicompendium.54202#sec-14> [Accessed: 30-09-2025].
- CABI, 2025. CABI Compendium [Web page]. Available online: <https://www.cabidigitallibrary.org/journal/cabicompendium> [Accessed: April 2025].
- Calari A, Paltrinieri S, Contaldo N, Sakalieva D, Mori N, Duduk B & Bertaccini A, 2011. Molecular evidence of phytoplasmas in winter oilseed rape, tomato and corn seedlings. *Bulletin of Insectology*, 64 (Supplement), S157-S158.
- Cameron H & Thompson M, 1985. Seed transmission of apple mosaic virus in hazelnut. *Proceedings of the XIII International Symposium on Fruit Tree Virus Diseases* 193, 131-132 pp.
- Caubel G, 1983. Epidemiology and control of seed-borne nematodes. *Seed Science and Technology*, International Seed Testing Association Ista, 11, 989-996.
- Chalanska A, Labanowski G & Malewski T, 2011. Detection of *Aphelenchoides ritzemabosi* (Schwartz) in scarlet sage (*Salvia splendens*) seeds. *Progress in Plant Protection*, 51 (3), 1095-1099. Available online: <http://www.progress.plantprotection.pl/content/view/170/14/lang,en/>
- Chigoziri E & Ekefan EJ, 2013. Seed borne fungi of chilli pepper (*Capsicum frutescens*) from pepper producing areas of Benue State, Nigeria. *Agriculture and Biology Journal of North America*, 4 (4), 370-374. Available online: <https://doi.org/10.5251/abjna.2013.4.4.370.374>
- Choi Y-J, Shin H-D & Thines M, 2009. Two novel *Peronospora* species are associated with recent reports of downy mildew on sages. *Mycological Research*, 113 (12), 1340-1350.
- Christen AA, 1982. Demonstration of *Verticillium-Albo-Atrum* within Alfalfa Seed. *Phytopathology*, 72 (4), 412-414.
- Collins B, 2016. Management of *Fusarium* wilt in bunching spinach production in Ontario, Canada. University of Guelph. Available online: URL
- Converse R, 1990. Virus and virus-like diseases of strawberry. *Hortscience* 25, 883-884.
- Converse RH, 1973. Occurrence and some properties of raspberry bushy dwarf virus in *Rubus* species in the United States. *Phytopathology*, 63 (6), 780.
- Converse RH & Stace-Smith R, 1987. Tomato ringspot virus in strawberry. . In: Converse R (ed.), *Virus Diseases of small Fruits*. Agriculture Handbook no. 631 United States Department of Agriculture, pp. 52-54.
- Credi R, Shier JL & Stace-Smith R, 1986. Occurrence of raspberry bushy dwarf virus in native thimbleberry in British Columbia. *Acta Horticulturae*, 186, 17-22. Available online: <https://doi.org/10.17660/ActaHortic.1986.186.2>
- Davalos N, Wells H & Ali A, 2021. Preliminary study of viruses infecting strawberry (*Fragaria* spp.) in the Midsouthern United States. *Plant Health Progress*, 23 (1), 7-13.
- Debode J, Van Hemelrijck W, Heungens K, Maes M & Creemers P, 2011. First report of *Pilidium concavum* causing tan-brown rot on strawberry fruit in Belgium. *Plant Disease*, 95 (8), 1029-1029.
- Delfosse VC, Barrios Barón MP & Distéfano AJ, 2021. What we know about poleroviruses: Advances in understanding the functions of polerovirus proteins. *Plant Pathology*, 70 (5), 1047-1061. Available online: <https://doi.org/10.1111/ppa.13368>
- Derbyshire DM, 1960. Seed transmission of *Didymella lycopersici*. *Plant Pathology*, 9 (4).
- Dey P, Hossain I, Mahmud H & Hossain MD, 2024. Management of seed borne *Ralstonia solanacearum* in Brinjal (*Solanum melongena* L.) and Tomato (*Lycopersicon esculentum* Mill.). *Bangladesh Journal of Botany*, 53 (1), 173-183. Available online: <https://doi.org/10.3329/bjb.v53i1.72279>
- Ding X, Chen D, Du Z, Zhang J & Wu Z, 2019. The complete genome sequence of a novel cytorhabdovirus identified in strawberry (*Fragaria ananassa* Duch.). *Archives of Virology*, 164 (12), 3127-3131.

- dos Santos e Silva MSB, Rodrigues AAC, Candido e Silva EK, de Oliveira ACS, de Oliveira LdJMG, Costa NdJF, Silva MRM & de Lemos RNS, 2019. Health quality and reduction of pathogenic transmission in tomato seeds using plant extracts. *Australian Journal of Crop Science*, 13 (4), 635-641. Available online: <https://doi.org/10.21475/ajcs.19.13.04.p1680>
- Douglas Gubler W & Converse RH, 1993. Diseases of strawberry (*Fragaria x ananassa* Duch.) [Web page]. American Phytopathological Society, St Paul, Minnesota. Available online: <https://www.apsnet.org/edcenter/resources/commonnames/Pages/Strawberry.aspx> [Accessed: 16-06-2025].
- EFSA-Panel-on-Plant-Health, 2013. Scientific opinion on the risk to plant health posed by Arabis mosaic virus, Raspberry ringspot virus, Strawberry latent ringspot virus and Tomato black ring virus to the EU territory with the identification and evaluation of risk reduction options. *EFSA Journal*, 11 (10), 3377-n/a. Available online: <https://doi.org/10.2903/j.efsa.2013.3377>
- EFSA Panel on Plant Health, Bragard C, Dehnen-Schmutz K, Di Serio F, Gonthier P, Jaques Miret JA, Justesen AF, MacLeod A, Magnusson CS, Milonas P, Navas-Cortes JA, Parnell S, Potting R, Reignault PL, Thulke H-H, Van der Werf W, Vicent Civera A, Yuen J, Zappalà L, Van der Wolf J, Kaluski T, Pautasso M & Jacques M-A, 2019. Pest categorisation of the *Ralstonia solanacearum* species complex. *EFSA Journal*, 17 (2), e05618. Available online: <https://doi.org/10.2903/j.efsa.2019.5618>
- EFSA Panel on Plant Health, Bragard C, Dehnen-Schmutz K, Gonthier P, Jacques M-A, Jaques Miret JA, Justesen AF, MacLeod A, Magnusson CS, Milonas P, Navas-Cortes JA, Parnell S, Potting R, Reignault PL, Thulke H-H, Van der Werf W, Vicent Civera A, Yuen J, Zappalà L, Candresse T, Chatzivassiliou E, Finelli F, Winter S, Bosco D, Chiumenti M, Di Serio F, Kaluski T, Minafra A & Rubino L, 2019a. Pest categorisation of non-EU viruses of *Fragaria* L. *EFSA Journal*, 17 (9), e05766. Available online: <https://doi.org/10.2903/j.efsa.2019.5766>
- EFSA Panel on Plant Health, Bragard C, Dehnen-Schmutz K, Gonthier P, Jacques M-A, Jaques Miret JA, Justesen AF, MacLeod A, Magnusson CS, Milonas P, Navas-Cortes JA, Parnell S, Potting R, Reignault PL, Thulke H-H, Werf Wvd, Vicent Civera A, Yuen J, Zappalà L, Candresse T, Chatzivassiliou E, Winter S, Chiumenti M, Di Serio F, Kaluski T, Minafra A & Rubino L, 2019b. List of non-EU viruses and viroids of *Cydonia* Mill., *Fragaria* L., *Malus* Mill., *Prunus* L., *Pyrus* L., *Ribes* L., *Rubus* L. and *Vitis* L. *EFSA Journal*, 17 (9), e05501. Available online: <https://doi.org/10.2903/j.efsa.2019.5501>
- El-Ammari S, Sariah M, Zainal Abidin M & Hor Y, 1997. Site of infection and transmission of *Collectotrichum gloeosporioides* in *Capsicum annum*. *Malaysian Applied Biology*, 26, 43-50.
- El-Nagerabi SAF, Elshafie AE & Abdalla AH, 2000. Composition of mycoflora and aflatoxins in pea seeds from the Sudan. *Kuwait Journal of Science & Engineering*, 27 (1), 109-121. Available online: <https://ovidsp.ovid.com/ovidweb.cgi?T=JS&CSC=Y&NEWS=N&PAGE=fulltext&D=bioba19&AN=BACD200000261953>
- EPPO, 2015. EPPO Global Database [Web page]. Available online: <https://gd.eppo.int> [Accessed: 01-04-2025].
- EPPO, 2021. Neopestalotiopsis species are causing emerging strawberry diseases worldwide. *EPPO Reporting Service*, 2021/229.
- EPPO, 2025a. Nepovirus nicotianae [Web page]. Available online: <https://gd.eppo.int/taxon/TRSV00/hosts> [Accessed: 30-09-2025].
- EPPO, 2025b. EPPO Global Database [Web page]. Available online: <https://gd.eppo.int> [Accessed: April - May 2025].
- Esquibet M, Bekal S, Castagnone-Sereno P, Gauthier J-P, Rivoal R & Caubel G, 1998. Differentiation of normal and giant *Vicia faba* populations of the stem nematode *Ditylenchus dipsaci*: agreement between RAPD and phenotypic characteristics. *Heredity*, 81 (3), 291-298.
- FAO, 2016. ISPM (International Standards for Phytosanitary Measures) No. 27. Diagnostic protocols for regulated pests. DP 17: *Aphelenchoides besseyi*, *A. fragariae* and *A. ritzemabosi*. Adopted 2016. FAO, Rome.
- FAO, 2021. ISPM (International standards for phytosanitary measures) No. 38. International movement of seeds. Food and Agricultural Organization of the United Nations, Secretariat of the International Plant Protection Convention. Available online: <https://www.ippc.int/en/core-activities/standards-setting/ispms/>
- Favoreto L, Santos JM, Calzavara SA & Lara LA, 2011. Phytosanitary study, multiplication and taxonomy of nematodes associated with seeds of forage grasses in Brazil. *Nematologia Brasileira*, 35 (1-2), 20-35.

- Ferrante P & Scortichini M, 2018. *Xanthomonas arboricola* pv. *fragariae*: a confirmation of the pathogenicity of the pathotype strain. *European Journal of Plant Pathology*, 150 (3), 825-829.
- Fowkes A, Adams IP, Jones RA, Fox A, McGreig S & Boonham N, 2022. Historical and recent tomato black ring virus and beet ringspot virus isolate genomes reveal interspecies recombination and plant health regulation inconsistencies. *Plant Pathology*, 71 (3), 729-740.
- Fránová-Honetšlegrová J, Erbenova M & Martin R, 1998. Isolation of tobacco necrosis virus from strawberry leaves in the Czech Republic. *Acta Virol*, 42, 325-331.
- Fránová-Honetšlegrová J, Mráz I, Nebesarova J & Sip M, 1999. Preferential banding of secondary veins in strawberry is caused by mixed virus infection. *Acta Virologica*, 43 (6), 349-355.
- Fránová J, Přebilová J & Koloniuk I, 2019. Molecular and biological characterization of a new strawberry cytorhabdovirus. *Viruses*, 11 (11), 982.
- Fuchs M, Bar-Joseph M, Candresse T, Maree HJ, Martelli GP, Melzer MJ, Menzel W, Minafra A, Sabanadzovic S & Report Consortium I, 2020. ICTV virus taxonomy profile: Closteroviridae. *Journal of General Virology*, 101 (4), 364-365.
- Fungorum I, 2025. *Stagonospora fragariae* Briard & Har. [Web page]. Available online: <https://www.indexfungorum.org/names/NamesRecord.asp?RecordID=225689> [Accessed: 23-05-2025].
- Garrido C, Carbú M, Fernández-Acero FJ, González-Rodríguez VE & Cantoral JM, 2011. New insights in the study of strawberry fungal pathogens. *Genes Genomes Genomics*, 5 (1), 24-39.
- Garrido C, González-Rodríguez VE, Carbú M, Husaini AM & Cantoral JM, 2016. Fungal diseases of strawberry and their diagnosis. In, *Strawberry: Growth, development and diseases*. CABI Wallingford UK, pp. 157-195.
- Gibbs A & Harrison B, 1964. Nematode-transmitted viruses in sugar beet in East Anglia. *Plant Pathology*, 13 (4).
- Gilbert RG & Peadar RP, 1988. Dissemination of *Verticillium-Albo-Atrum* in Alfalfa by Internal Seed Inoculum. *Canadian Journal of Plant Pathology*, 10 (1), 73-77.
- Golino D, Cunningham M, Rowhani A & Sim S, 2005. Transmission of rose mosaic viruses. *Proceedings of the IV International Symposium on Rose Research and Cultivation* 751, 217-224 pp.
- Golnaraghi A, Shahraeen N, Pourrahim R, Farzadfar S & Ghasemi A, 2004. Occurrence and relative incidence of viruses infecting soybeans in Iran. *Plant Disease*, 88 (10), 1069-1074.
- Grove GG, Eastwell KC, Jones AL & Sutton TB, 2003. Diseases of apple. In, *Apples: botany, production and uses*. CABI Publishing Wallingford UK, pp. 459-488.
- Guimarães GR, Pereira FS, Matos FS, Mello S & Carvalho DD, 2014. Suppression of seed borne *Cladosporium herbarum* on common bean seed by *Trichoderma harzianum* and promotion of seedling development. *Tropical Plant Pathology*, 39, 401-406.
- Gullino ML, Gilardi G & Garibaldi A, 2012. More from seed borne pathogens in leafy vegetables. *Informatore Agrario*, 68 (37), 60-64. Available online: <http://www.informatoreagrario.it>
- Hajihassani A, Tenuta M & Gulden RH, 2016. Host preference and seedborne transmission of *Ditylenchus weischeri* and *D. dipsaci* on select pulse and non-pulse crops grown in the Canadian Prairies. *Plant Disease*, 100 (6), 1087-1092.
- Hajizadeh M, Zandan NG, Koloniuk I, Sierra-Mejia A & Tzanetakis IE, 2025. Characterization, Detection, and Prevalence of a Novel Strawberry Crinivirus. *Plant Disease*, 109 (5), 988-991. Available online: <https://doi.org/10.1094/pdis-08-24-1682-sc>
- Harrison B, 1957. Studies on the host range, properties and mode of transmission of beet ringspot virus. *Annals of Applied Biology*, 45 (3), 462-472. Available online: <https://doi.org/10.1111/j.1744-7348.1957.tb05883.x>
- Hayden N & Maude R, 1992. The role of seed-borne *Aspergillus niger* in transmission of black mould of onion. *Plant Pathology*, 41 (5), 573-581. Available online: <https://doi.org/10.1111/j.1365-3059.1992.tb02456.x>
- He C, Zhao X, Fan L, Li S & Wang H, 2022. Strawberry, a new natural host of brassica yellows virus in China. *Plant Disease*, 106 (3).
- Hernandez-Perez P, 2005. Management of seedborne *Stemphylium botryosum* and *Cladosporium variabile* causing leaf spot of spinach seed crops in western Washington. Washington State University Pullman, WA. Available nline: URL
- Heydari N, Ghorbani M, Salari M, Panjehkeh N & Pirnia M, 2017. New records of anamorphic fungi from North of Iran. *Mycologia Iranica*, 4 (1), 49-59.
- Huang Q-r, Niu Y-h, Wu K, Liu Z-j, Cao M, Zhao L & Wu Y-f, 2021. Lncrna sequencing identification of viruses infecting strawberry in Shaanxi. *Acta Horticulturae Sinica* November, 48 (8), 1589-1594. Available online: <https://doi.org/10.16420/j.issn.0513-353x.2020-0675>
- ICTV, 2025a. Species List: Solemoviridae [Web page]. International Committee on Taxonomy of Viruses. Available online:

- <https://ictv.global/report/chapter/solemoviridae/taxonomy/solemoviridae> [Accessed: 09-12-2025].
- ICTV, 2025b. International Committee on Taxonomy of Viruses (ICTV) [Web page]. Available online: <https://ictv.global/taxonomy/> [Accessed: 01-10-2025].
- ICTV, 2026a. Genus: Crinivirus [Web page]. International Committee on Taxonomy of Viruses. Available online: <https://ictv.global/report/chapter/closteroviridae/closteroviridae/crinivirus> [Accessed: 25-03-2026].
- ICTV, 2026b. Genus: Caulimovirus [Web page]. International Committee on Taxonomy of Viruses. Available online: <https://ictv.global/report/chapter/caulimoviridae/caulimoviridae/caulimovirus> [Accessed: 25-03-2026].
- ICTV, 2026c. Genus: Nepovirus [Web page]. International Committee on Taxonomy of Viruses. Available online: <https://ictv.global/report/chapter/secoviridae/secoviridae/nepovirus> [Accessed: 25-03-2025].
- Ji S, Li H, Zhou Y, Li X, Yan J & Zhang W, 2023. First report of bacterial wilt caused by *Enterobacter mori* of strawberry in Beijing, China. *Plant Disease*, 107 (6), 1936.
- Johnson H, Converse R, Amorao A, Espejo J & Frazier N, 1984. Seed transmission of tobacco streak virus in strawberry. *Plant Disease*, 68, 390-392.
- Jones RA, Fowkes AR, McGreig S, Fox A & Adams IP, 2022. Forty-year-old beet ringspot virus isolate: tests for potato true seed transmission, genome sequencing, recombination analysis and phylogenetic placement. *Journal of Plant Pathology*, 106 (2), 389-393.
- Jordan R, Molloy D, Guaragna M & Lockhart B, 2019. Detection and first report of beet ringspot virus in ornamental *Oxalis* in the United States. *Plant Disease*, 103 (7), 1800.
- Kacharmazov V & Khristov L, 1976. Studies on the causes of decreased viability of strawberry hybrids. 1. The appearance and spread of virus infection in breeding nurseries.
- Kaiser WJ, Wyatt S & Klein R, 1991. Epidemiology and seed transmission of two tobacco streak virus pathotypes associated with seed increases of legume germ plasm in eastern Washington.
- Kgatle M, Truter M, Ramusi T, Flett B & Aveling T, 2018. *Alternaria alternata*, the causal agent of leaf blight of sunflower in South Africa. *European Journal of Plant Pathology*, 151, 677-688.
- Khatun A, Shamsi S & Bashar M, 2023. Transmission of pathogenic fungi from seeds to seedlings in cotton (*Gossypium hirsutum* L.). *Dhaka University Journal of Biological Sciences*, 32 (2), 225-232. Available online: <https://doi.org/10.3329/dujbs.v32i2.67681>
- Kirdat K, Tiwarekar B, Swetha P, Padma S, Thorat V, Manjula KN, Kavya N, Sundararaj R & Yadav A, 2022. Nested real-time PCR assessment of vertical transmission of sandalwood spike phytoplasma (*Ca. phytoplasma asteris*). *Biology*, 11 (10), 1494.
- Kis S, Salamon P, Kis V & Szittya G, 2017. Molecular characterization of a beet ringspot nepovirus isolated from *Begonia ricinifolia* in Hungary. *Archives of Virology*, 162 (11), 3559-3562.
- Koloniuk I, Přebyllová J, Čmejla R, Valentová L & Fránová J, 2022. Identification and characterization of a novel umbra-like virus, strawberry virus A, infecting strawberry plants. *Plants*, 11 (5), 643.
- Koycu ND & Ozer N, 1997. Determination of seedborne fungi in onion and their transmission to onion sets. *Phytoparasitica*, 25 (1), 25-31.
- Kumar R & Gupta A, 2020. Seed-borne diseases of agricultural crops: detection, diagnosis & management. Springer.
- Lacey M, 1939. Studies on a bacterium associated with leafy galls, fasciations and "cauliflower" disease of various plants. Part III. Further isolations, inoculation experiments and cultural studies. *Ann Appl Biol*, 26, 262-278.
- Lee D-H, 1995. Seed-borne infection of anthracnose fungi isolated from diseased red pepper. *The Korean Journal of Mycology*, 23 (2), 114-120.
- Lenz O, Přebyllová J, Fránová J & Koloniuk I, 2020. *Fragaria vesca*-associated virus 1: A new virus related to negeviruses. *Archives of Virology*, 165 (5), 1249-1252.
- Lister R & Murant A, 1967. Seed-transmission of nematode-borne viruses. *Annals of Applied Biology*, 59 (1), 49-62.
- Lopes U, Zambolim L, Lopes U, Pereira O & Costa H, 2010. First report of *Pilidium concavum* causing tan-brown rot in strawberry fruits in Brazil.
- Lowe-Power T, 2025. *Ralstonia pseudosolanacearum* [Web page].
- Luigi M, Faggioli F, Barba M & Giunchedi L, 2010. The molecular characterization of HSVd isolates associated with dapple fruit and fruit rugosity in plum seedlings suggests a possible role of breeding in viroid dissemination. *Julius-Kühn-Archiv*, (427), 101.
- Lunn J, 1977. *Rhizopus sexualis*. CMI Descriptions of Pathogenic Fungi and Bacteria. CAB International.

- Machmud M & Middleton KJ, 1990. Seed infection and transmission of *Pseudomonas solanacearum* on groundnut. ACIAR Proceedings Series, 31, 57.
- Marin MV, Carvalho R, Paret ML, Jones JB & Peres NA, 2024. *Pseudomonas fragariae* sp. nov., a novel bacterial species causing leaf spots on strawberry (*Fragaria × ananassa*). International journal of systematic and evolutionary microbiology, 74 (8), 006476.
- Martin R, 2001. Virus diseases of Rubus and strategies for their control. Acta Horticulturae, 585, 265-270.
- McGuire JM, 1973. Retention of tobacco ringspot virus by *Xiphinema americanum*. Phytopathology, 63 (2), 324-326. Available online: <https://doi.org/10.1094/Phyto-63-324>
- Medberry A & Tzanetakis IE, 2022. Identification, characterization, and detection of a novel strawberry cytorhabdovirus. Plant Disease, 106 (11), 2784-2787. Available online: <https://doi.org/10.1094/pdis-11-21-2449-sc>
- Medberry AN, Srivastava A, Diaz-Lara A, Rwahnih MA, Villamor DE & Tzanetakis IE, 2023. A novel, divergent member of the Rhabdoviridae family infects strawberry. Plant Disease, 107 (3), 620-623.
- Mellor FC & Stace-Smith R, 1963. Reaction of strawberry to a ringspot virus from raspberry. Canadian journal of botany, 41 (6), 865-870.
- Mink GI, 1993. Pollen and Seed-Transmitted Viruses and Viroids. Annual review of phytopathology, 31 (Volume 31, 1993), 375-402. Available online: <https://doi.org/10.1146/annurev.py.31.090193.002111>
- Möller S & Maruthi M, 2025. A review of factors affecting the success of geminivirus infectious clones. Plant Cell Reports, 44 (8), 189.
- Motsar E, Sheveleva A, Sharko F, Petrova K, Slobodova N, Murataev R, Mitrofanova I & Chirkov S, 2025. Molecular characterization of tobacco streak virus, beet ringspot virus, and beet ringspot virus satellite RNA from a new natural host, *Phlox paniculata*. Plants, 14 (11), 1619.
- Mountain WL, Powell C, Forer L & Stouffer R, 1983. Transmission of tomato ringspot virus from dandelion via seed and dagger nematodes.
- Murant A, 1983. Seed and pollen transmission of nematode-borne viruses. Seed Science and Technology (Netherlands), 11 (3).
- Murant A, Chambers J & Jones A, 1974. Spread of raspberry bushy dwarf virus by pollination, its association with crumbly fruit, and problems of control. Annals of Applied Biology, 77 (3), 271-281.
- MycoBank, 2025. MycoBank Database [Web page]. Available online: <https://www.mycobank.org/> [Accessed: April 2025].
- Naktuinbouw, 2024. Aardbei uit zaad in de lift. Buitenstebinnen, 22, 16-17.
- NBNAtlas, 2025. *Stagonospora fragariae* Briard & Har. [Web page]. Available online: <https://species.nbnatlas.org/species/BMSSYS0000017861> [Accessed: 23-05-2025].
- NVWA, 2025. Register Q-organismen, v. 4.05, 11/03/2025 [Web page]. Netherlands Food and Consumer Product Safety Authority. Available online: <https://www.nvwa.nl/documenten/export/fytsanitair/voorschriften/algemeen/register-q-organismen-en-q-waardige-organismen> [Accessed: 01-04-2025].
- Ormerod P, 1970. A virus associated with Loganberry degeneration disease.
- Ozgonen H & Gulcu M, 2011. Determination of mycoflora of pea (*Pisum sativum*) seeds and the effects of *Rhizobium leguminosorum* on fungal pathogens of peas. African Journal of Biotechnology, 10 (33), 6235-6240.
- Öztürk M & Soyly S, 2022. A new disease of strawberry, bacterial blight caused by *Erwinia amylovora* in Turkey. Journal of Plant Pathology, 104 (1), 269-280.
- Pachoute J, dos Santos GR & de Souza DJ, 2024. Antagonistic effects of *Beauveria bassiana* on seed-borne fungi of cowpea (*Vigna unguiculata*). Biologia (Bratislava), 79 (5), 1487-1495. Available online: <https://doi.org/10.1007/s11756-024-01615-7>
- Parsa S, García-Lemos AM, Castillo K, Ortiz V, López-Lavalle LAB, Braun J & Vega FE, 2016. Fungal endophytes in germinated seeds of the common bean, *Phaseolus vulgaris*. Fungal Biology, 120 (5), 783-790.
- Parte AC, Sardà Carbasse J, Meier-Kolthoff JP, Reimer LC & Göker M, 2020. List of Prokaryotic names with Standing in Nomenclature (LPSN) moves to the DSMZ. International journal of systematic and evolutionary microbiology, 70 (11), 5607-5612. Available online: <https://doi.org/10.1099/ijsem.0.004332>
- Perello AE & Larran S, 2013. Nature and effect of *Alternaria* spp. complex from wheat grain on germination and disease transmission. Pakistan Journal of Botany, 45 (5), 1817-1824.
- Petkar A & Ji P, 2017. Infection courts in watermelon plants leading to seed infestation by *Fusarium oxysporum* f. sp. *niveum*. Phytopathology, 107 (7), 828-833. Available online: <https://doi.org/10.1094/phyto-12-16-0429-r>

- Phillips D, 1956. Tomato seed transmission of *Didymella lycopersici* Kleb. Transactions of the British Mycological Society, 39 (3), 319-329.
- PlantAtlas, 2025. Digital Plant Atlas [Web page]. Rijksuniversiteit Groningen & Deutsches Archäologisches Institut - Berlin. Available online: <https://www.plantatlas.eu/> [Accessed: 22-05-2025].
- Postman JD & Mehlenbacher SA, 1992. Apple mosaic virus in hazelnut germplasm. Proceedings of the III International Congress on Hazelnut 351, 601-610 pp.
- Preeti Aggarwal PA & Sood AK, 2005. Detection of seed-borne infection of bacterial wilt in *Capsicum*. In: Sharma RC & Sharma JN (eds.). Scientific Publishers (India), Jodhpur.
- Pringle C, 1998. Virus taxonomy-San Diego 1998. Archives of Virology, 143 (7), 1449.
- Pscheidt J & Ocamb C, 2025. Strawberry (*Fragaria* spp.)-Nematode, stem and bulb [Web page]. Available online: <https://pnwhandbooks.org/node/3567> [Accessed: 26-05-2025].
- Putnam ML & Miller ML, 2007. *Rhodococcus fascians* in Herbaceous Perennials. Plant Disease, 91 (9), 1064-1076. Available online: <https://doi.org/10.1094/pdis-91-9-1064>
- Reeder R, 2020. *Diplodia seriata* (grapevine trunk disease) [Web page]. Available online: <https://www.cabidigitallibrary.org/doi/full/10.1079/cabicompendium.9630#sec-16> [Accessed: 23-05-2025].
- Richardson MJ, 1990. An annotated list of seed-borne diseases. Department of Agriculture and Fisheries or Scotland, Edinburgh.
- Richter U & Jage H, 2003. Noteworthy records of phytoparasitic microfungi from Germany and Austria: looking back on Werner Lehmann's 80th birthday. Boletus, 26 (2), 127-130.
- Rijavec T, Lapanje A, Dermastia M & Rupnik M, 2007. Isolation of bacterial endophytes from germinated maize kernels. Canadian Journal of Microbiology, 53 (6), 802-808.
- Rivera A, Morales-Mora L, Mauricio-Gutiérrez A, Romero-Arenas O, Contreras-Paredes C & Villar-Ruano N, 2024. New fungal disease of strawberry fruits caused by *Neurospora dictyophora* in Mexico. New Disease Reports, 49 (1), e12261.
- Roberts P, Jones J, Chandler C, Stall R & Berger R, 1996. Survival of *Xanthomonas fragariae* on strawberry in summer nurseries in Florida detected by specific primers and nested polymerase chain reaction.
- Rose C & Damm U, 2024. Diversity of *Colletotrichum* species on strawberry (*Fragaria* × *ananassa*) in Germany. Phytopathologia Mediterranea, 63 (2), 155-178.
- Sackston W & Martens J, 1959. Dissemination of *Verticillium albo-atrum* on seed of sunflower (*Helianthus annuus*). Canadian journal of botany, 37 (5), 759-768.
- Safni I, Cleenwerck I, De Vos P, Fegan M, Sly L & Kappler U, 2014. Polyphasic taxonomic revision of the *Ralstonia solanacearum* species complex: proposal to emend the descriptions of *Ralstonia solanacearum* and *Ralstonia syzygii* and reclassify current *R. syzygii* strains as *Ralstonia syzygii* subsp. *syzygii* subsp. nov., *R. solanacearum* phylotype IV strains as *Ralstonia syzygii* subsp. *indonesiensis* subsp. nov., banana blood disease bacterium strains as *Ralstonia syzygii* subsp. *celesbesensis* subsp. nov. and *R. solanacearum* phylotype I and III strains as *Ralstonia pseudosolanacearum* sp. nov. International journal of systematic and evolutionary microbiology, 64 (9), 3087-3103.
- Saharan GS & Mehta N, 2008. Sclerotinia diseases of crop plants: biology, ecology and disease management. Springer Science & Business Media.
- Saponari M, Savino V & Martelli G, 2002. Seed transmission in olive of two olive-infecting viruses.
- Sastry KS, 2013. Seed-borne plant virus diseases. Springer science & business media.
- Sastry KS, Mandal B, Hammond J, Scott SW & Briddon RW, 2019. Encyclopedia of plant viruses and viroids. Springer.
- Schimanski H, 1987. Investigations concerning seed transmissibility of tomato black ring virus in black locust (*Robinia pseudoacacia* L.) and European elder (*Sambucus nigra* L.). Zentralblatt für Mikrobiologie, 142 (6), 477-481.
- Sharman M, Thomas J & Persley DM, 2015. Natural host range, thrips and seed transmission of distinct tobacco streak virus strains in Queensland, Australia. Annals of Applied Biology, 167 (2), 197-207.
- Song P, Li G, Zhao Q, Lu G, Zhao X, Liu L, Song Y, Niu J & Zhou H, 2023. First report of a new bacterial stem rot disease of strawberry (*Fragaria* × *ananassa*) caused by *Pantoea ananatis* in Jiangsu, China. Plant Disease, 107 (7), 2210.
- Spiegel S, Martin RR, Leggett F, Borg Mt & Postman J, 1993. Characterization and geographical distribution of a new ilarvirus from *Fragaria chiloensis*. Phytopathology, 83 (9), 991-995. Available online: <https://doi.org/10.1094/Phyto-83-991>
- Štefanac Z & Wrischer M, 1983. Spinach latent virus: some properties and comparison of two isolates. Acta Botanica Croatica, 42 (1), 1-9.

- Subbotin SA, 2024. Rapid detection of the strawberry foliar nematode *Aphelenchoides fragariae* using recombinase polymerase amplification assay with lateral flow dipsticks. *International Journal of Molecular Sciences*, 25 (2), 844.
- Sumithra KU, Krishnappa M, Vasanth TK, Shetty HS, Mortensen CN & Mathur SB, 2000. Seed-borne nature of *Ralstonia solanacearum* in eggplant (*Solanum melongena* L.) cultivars in India. *Seed Science & Technology*, 28 (2), 291-299.
- Tilford PE, 1936. Fasciation of sweet potatoes caused by *Phytomonas fascians* n.sp. *Journal of Agricultural Research*, 53, 383.
- Tu J, 1983. Epidemiology of anthracnose caused by *Colletotrichum lindemuthianum* on white bean (*Phaseolus vulgaris*) in southern Ontario: survival of the pathogen. *Plant Disease* 67, 402-404.
- Tzanetakis I, Aknadibossian V, Špak J, Constable F, Harper S, Hammond J, Candresse T, Folimonova S, Freitas-Astúa J, Fuchs M & et al., 2024. Streamlining global germplasm exchange: integrating scientific rigor and common sense to exclude phantom agents from regulation. *Plant Disease*, PDIS-04-24-0745-FE.
- Tzanetakis I, Halgren A, Wintermantel W, Keller K & Martin R, 2004a. Two criniviruses are associated with the strawberry pallidosis disease. *Acta Horticulturae*, 656, 21-26.
- Tzanetakis I, Mackey I & Martin R, 2004b. Strawberry necrotic shock virus is a distinct virus and not a strain of Tobacco streak virus. *Archives of Virology*, 149, 2001-2011.
- Tzanetakis IE, 2012. Tobacco streak virus (tobacco streak) [Web page]. Available online: <https://www.cabidigitallibrary.org/doi/full/10.1079/cabicompendium.48107#sec-12>
- Tzanetakis IE & Martin RR, 2013. Expanding field of strawberry viruses which are important in North America. *International Journal of Fruit Science*, 13 (1-2), 184-195.
- Tzanetakis IE, Price R & Martin RR, 2008. Nucleotide sequence of the tripartite *Fragaria chiloensis* cryptic virus and presence of the virus in the Americas. *Virus Genes*, 36 (1), 267-272.
- Umesha S, Kavitha R & Shetty HS, 2005. Transmission of seed-borne infection of chilli by *Burkholderia solanacearum* and effect of biological seed treatment on disease incidence. *Archives of Phytopathology and Plant Protection*, 38 (4), 281-293. Available online: <https://doi.org/10.1080/03235400500094209>
- USDA, 2025. USDA Fungal Databases [Web page]. Available online: <https://fungi.ars.usda.gov/> [Accessed: 13-06-2025].
- Van Hemelrijck W, Ceustermans A, Van Campenhout J, Lieten P & Bylemans D, 2016. Crown rot in strawberry caused by *Pestalotiopsis*. *Acta Horticulturae*, 1156, 781-786.
- Villamor D, Keller K, Martin R & Tzanetakis I, 2022. Comparison of high throughput sequencing to standard protocols for virus detection in berry crops. *Plant Disease*, 106 (2), 518-525.
- Walcott RR, Gitaitis RD, Castro AC, Sanders FH, Jr. & Diaz-Perez JC, 2002. Natural infestation of onion seed by *Pantoea ananatis*, causal agent of center rot. *Plant Disease*, 86 (2), 106-111. Available online: <https://doi.org/10.1094/PDIS.2002.86.2.106>
- Walker PJ, Bejerman N, Blasdell KR, Debat H, Dietzgen RG, Fooks AR, Freitas-Astúa J, Garver K, Kondo H, Ramos-González P, Shi M, Tesh R, Tordo N, Vasilakis N & Whitfield AE, 2026. Family: Rhabdoviridae [Web page]. Available online: <https://ictv.global/report/chapter/rhabdoviridae/rhabdoviridae> [Accessed: 24-03-2026].
- Wan Chow Wah Y & Symons R, 1999. Transmission of viroids via grape seeds. *Journal of Phytopathology*, 147 (5), 285-291.
- Waterhouse G & Waterston J, 1964. *Phytophthora nicotianae* var. *nicotianae*. CMI Descriptions of Pathogenic Fungi and Bacteria, (4): Sheet 33.
- Wei W & Zhao Y, 2025. Phytoplasma transmission via seeds: a hypothesis awaiting definitive evidence. In, *Phytoplasmas: Genomes, Plant Hosts and Vectors*. CABI GB, pp. 94-105.
- Wenneker M, 2022. *Erwinia pyrifoliae* (Asian pear blight). CABI Compendium, 27 (51264). Available online: <https://doi.org/10.1079/cabicompendium.51264>
- Wenneker M, Bergsma-Vlami M & Van Der Steen J, 2017. Epidemiology of *Erwinia pyrifoliae*, a new pathogen on strawberry in The Netherlands. *Acta Horticulturae*, 1156, 721-726.
- Yang Y, Kim KS & Anderson EJ, 1997. Seed transmission of cucumber mosaic virus in spinach. *Phytopathology*, 87 (9), 924-931. Available online: <https://doi.org/10.1094/phyto.1997.87.9.924>
- Zad SJ, 1992. Safflower seed-borne diseases. *Mededelingen van de Faculteit Landbouwwetenschappen, Universiteit Gent*, 57 (2a), 161-163.
- Zhang YX, Hua JY & He LY, 1993. Effect of infected groundnut seeds on transmission of *Pseudomonas solanacearum*. *Bacterial Wilt Newsletter*, 9, 9-10.
- Zhao L-l, Shi Z-j, Li T-t, Zhong J, Ding M & Zhang Z-k, 2020. Natural hosts and genetic diversity of *Crassocephalum* yellow vein virus. *Journal of Plant Protection*, 47 (3), 647-656.
- Zreik L, Bové JM & Garnier M, 1998. Phylogenetic characterization of the bacterium-like organism associated with marginal chlorosis of strawberry and proposition of a Candidates taxon for

the organism, '*Candidatus Phlomobacter fragariae*'. International journal of systematic and evolutionary microbiology, 48 (1), 257-261.

Annex A List of organisms included in the assessment

Bacteria

Current scientific name (1. EPPO Global Database, 2. LPSN)	Name organism in publication or legislation	EPPO code	Source
<i>Candidatus Phlomobacter fragariae</i>	<i>Candidatus Phlomobacter fragariae</i>	PHMBFR	(Zreik et al., 1998)
<i>Enterobacter mori</i>	<i>Enterobacter mori</i>	ENTBMO	(Ji et al., 2023)
<i>Erwinia amylovora</i>	<i>Erwinia amylovora</i>	ERWIAM	(Öztürk & Soyulu, 2022)
<i>Erwinia pyrifoliae</i>	<i>Erwinia pyrifoliae</i>	ERWIPY	(Wenneker et al., 2017)
<i>Pantoea ananatis</i>	<i>Pantoea ananatis</i>	ERWIAN	(Song et al., 2023)
<i>Pseudomonas fragariae</i>	<i>Pseudomonas fragariae</i>	-	(Marin et al., 2024)
<i>Ralstonia pseudosolanacearum</i>	<i>Ralstonia pseudosolanacearum</i>	RALSPS	(Lowe-Power, 2025)
<i>Rhodococcus fascians</i>	<i>Rhodococcus fascians</i> (<i>Corynebacterium fascians</i> in publication)	CORBFA	(Bradbury, 1967)
<i>Xanthomonas arboricola</i> <i>pv. fragariae</i>	<i>Xanthomonas arboricola</i> <i>pv.</i> <i>fragariae</i>	XANTAF	(Ferrante & Scortichini, 2018)
<i>Xanthomonas fragariae</i>	<i>Xanthomonas fragariae</i>	XANTFR	(Roberts et al., 1996)
<i>Xylella fastidiosa</i>	<i>Xylella fastidiosa</i>	XYLEFA	(Bradbury, 1991)

Fungi and oomycetes

(source: Garrido et al. (2016) unless stated otherwise)

Current scientific name (1. EPPO Global Database, 2. Mycobank)	Name organism in publication or legislation	EPPO code
<i>Alternaria alternata</i>	<i>Alternaria alternata</i>	ALTEAL
<i>Alternaria tenuissima</i>	<i>Alternaria tenuissima</i>	ALTETE
<i>Armillaria mellea</i>	<i>Armillaria mellea</i>	ARMIME
<i>Aspergillus niger</i>	<i>Aspergillus niger</i>	ASPENI
<i>Athelia rolfsii</i>	<i>Sclerotium rolfsii</i>	SCLORO
<i>Boeremia lycopersici</i>	<i>Phoma lycopersici</i>	DIDYLY
<i>Botrytis cinerea</i>	<i>Botrytis cinerea</i>	BOTRCI
<i>Cercospora fragariae</i>	<i>Cercospora fragariae</i>	not available
<i>Cladosporium</i>	<i>Cladosporium</i> <i>spp.</i>	1CLADG
<i>Colletotrichum fragariae</i>	<i>Colletotrichum fragariae</i>	COLLFR
<i>Colletotrichum acutatum sensu lato</i> ¹	<i>Colletotrichum acutatum sensu lato</i>	COLLAC
<i>Colletotrichum</i> <i>spp.</i> other than <i>C. acutatum</i> <i>sensu lato</i> and <i>C. fragariae</i>	See main text section 3.3	

Current scientific name (1. EPPO Global Database, 2. Mycobank)	Name organism in publication or legislation	EPPO code
<i>Coniella straminea</i>	<i>Schizoparme straminea</i>	SKHZST
<i>Coniothyrium fragariae</i>	<i>Coniothyrium fragariae</i>	CONLFR
<i>Cylindrocarpon destructans</i>	<i>Cylindrocarpon destructans</i>	NECTRA
<i>Dematophora necatrix</i>	<i>Rosellinia necatrix</i>	ROSLNE
<i>Diplocarpon fragariae</i>	<i>Diplocarpon earlianum</i>	DIPCEA
<i>Fusarium oxysporum</i>	<i>Fusarium oxysporum</i>	FUSAOX
<i>Fusarium sambucinum</i>	<i>Fusarium sambucinum</i>	GIBBPU
<i>Gnomoniopsis comari</i>	<i>Gnomonia comari</i>	GNMPCO
Identity not clear	<i>Sphaeropsis malorum</i>	not available
<i>Idriella lunata</i>	<i>Idriella lunata</i>	not available
<i>Macrophomina phaseolina</i>	<i>Macrophomina phaseolina</i>	MCPHPH
<i>Mucor hiemalis</i>	<i>Mucor hiemalis</i>	MUCOHI
<i>Mucor mucedo</i>	<i>Mucor mucedo</i>	MUCOMU
<i>Mucor piriformis</i>	<i>Mucor piriformis</i>	MUCOPI
<i>Mycosphaerella louisianae</i>	<i>Mycosphaerella louisianae</i>	not available
Not found in the databases	<i>Sclerotium fragariae</i>	not available
<i>Olpidium brassicae sensu lato</i>	<i>Olpidium brassicae</i>	OLPIBR
<i>Paraconiothyrium fuckelii</i>	<i>Coniothyrium fuckelii</i>	LEPTCO
<i>Paraphomopsis obscurans</i>	<i>Phomopsis obscurans</i>	PHOPOB
<i>Passalora vexans</i>	<i>Cercospora vexans</i>	not available
<i>Penicillium aurantiogriseum</i>	<i>Penicillium cyclopium</i>	PENIAU
<i>Penicillium expansum</i>	<i>Penicillium expansum</i>	PENIEX
<i>Penicillium glabrum</i>	<i>Penicillium frequentans</i>	PENIGB
<i>Penicillium purpurogenum</i>	<i>Penicillium purpurogenum</i>	PENIPU
<i>Pestalotiopsis longisetula</i>	<i>Pestalotia longisetula</i>	PESTLO
<i>Phytophthora bisheria</i>	<i>Phytophthora bisheria</i>	PHYTBS
<i>Phytophthora cactorum</i>	<i>Phytophthora cactorum</i>	PHYTCC
<i>Phytophthora citricola</i>	<i>Phytophthora citricola</i>	PHYTCI
<i>Phytophthora citrophthora</i>	<i>Phytophthora citrophthora</i>	PHYTCO
<i>Phytophthora cryptogea</i>	<i>Phytophthora cryptogea</i>	PHYTCR
<i>Phytophthora fragariae</i>	<i>Phytophthora fragariae</i>	PHYTFR
<i>Phytophthora megasperma</i>	<i>Phytophthora megasperma</i>	PHYTME
<i>Phytophthora nicotianae var. nicotianae</i>	<i>Phytophthora nicotianae</i>	PHYTNN
<i>Pilidium lythri</i>	<i>Hainesia lythri</i>	DISHOE
<i>Podosphaera aphanis</i> ¹	<i>Podosphaera aphanis</i>	PODOAP
<i>Pythium ultimum</i>	<i>Pythium ultimum</i>	PYTHUL

Current scientific name (1. EPPO Global Database, 2. Mycobank)	Name organism in publication or legislation	EPPO code
<i>Ramularia grevilleana</i>	<i>Mycosphaerella fragariae</i>	MYCOFR
<i>Rhizoctonia fragariae</i>	<i>Rhizoctonia fragariae</i>	RHIZFR
<i>Rhizoctonia solani</i>	<i>Rhizoctonia solani</i>	RHIZSO
<i>Rhizopus sexualis</i>	<i>Rhizopus sexualis</i>	RIZPSE
<i>Rhizopus stolonifer</i>	<i>Rhizopus stolonifer</i>	RIZPST
<i>Sclerotinia sclerotiorum</i>	<i>Sclerotinia sclerotiorum</i>	SCLESC
<i>Septoria aciculosa</i>	<i>Septoria aciculosa</i>	not available
<i>Septoria fragariae</i>	<i>Septoria fragariae</i>	SEPTFR
<i>Septoria fragariaecola</i>	<i>Septoria fragariaecola</i>	not available
<i>Setophoma terrestris</i>	<i>Phoma terrestris</i>	PYRETE
<i>Stagonospora fragariae</i>	<i>Stagonospora fragariae</i>	STAGFR
<i>Verticillium albo-atrum sensu lato</i>	<i>Verticillium albo-atrum</i>	VERTAA
<i>Verticillium dahliae</i>	<i>Verticillium dahliae</i>	VERTDA

¹ Source: see main text section 3.3

Viruses and viroids

Name in publication (acronym)	Binomial name	EPPO code	Source
Apple mosaic virus (ApMV)	<i>Iilarvirus ApMV</i>	APMV00	(EFSA Panel on Plant Health et al., 2019b)
Apple rootstock virus A (ApRVA)	-	not available	(Villamor et al., 2022)
Arabis mosaic virus (ArMV)	<i>Nepovirus arabis</i>	ARMV00	(EFSA Panel on Plant Health et al., 2019b)
Beet pseudoyellows virus (BPYV)	<i>Crinivirus pseudobetae</i>	BPYV00	(EFSA Panel on Plant Health et al., 2019b)
Beet ringspot virus (BRSV)	<i>Nepovirus betae</i>	BRSV00	(Lister & Murrant, 1967; Fowkes et al., 2022)
Crassocephalum yellow vein virus (CraYVV)	<i>Begomovirus crassocephali</i>	CRAYVV	(Zhao et al., 2020)
Cucumber mosaic virus (CMV)	<i>Cucumovirus CMV</i>	CMV000	(EFSA Panel on Plant Health et al., 2019b)
Fragaria chiloensis cryptic virus (FCCV)	-	FCLCV0	(EFSA Panel on Plant Health et al., 2019b)
Fragaria chiloensis latent virus (FCiLV)	<i>Iilarvirus FCiLV</i>	FCiLV0	(EFSA Panel on Plant Health et al., 2019b)
Fragaria vesca-associated virus 1 (FVaV-1) ¹	-	not available	(Lenz et al., 2020)

Name in publication (acronym)	Binomial name	EPPO code	Source
Hop stunt viroid (HSVd)	<i>Hostuviroid impedi humuli</i>	HSVD00	(EFSA Panel on Plant Health et al., 2019b)
Lycopersicon esculentum nepovirus (LENV)	-	not available	(Huang et al., 2021)
Olive latent virus 1	<i>Alphanecrovirus oleae</i>	OLV100	(Fránová et al., 2019)
Plum bark necrosis stem pitting-associated virus (PBNSPaV)	<i>Ampelovirus pruni</i>	PBNSPA	(EFSA Panel on Plant Health et al., 2019b)
Raspberry bushy dwarf virus (RBDV) ²	<i>Idaeovirus rubi</i>	RBDV00	(Davalos et al., 2021)
Raspberry ringspot virus (RpRSV)	<i>Nepovirus rubi</i>	RPRSV0	(EFSA Panel on Plant Health et al., 2019b)
Spinach latent virus (SpLV)	<i>Iilarvirus SLV</i>	SPLV00	(Huang et al., 2021)
Strawberry chlorotic fleck-associated virus (SCFaV)	<i>Closterovirus fragariae</i>	SCFAV0	(EFSA Panel on Plant Health et al., 2019b)
Strawberry crinivirus 3 (SCrV-3)	-	not available	(EFSA Panel on Plant Health et al., 2019b)
Strawberry crinivirus 4 (SCrV-4)	-	not available	(EFSA Panel on Plant Health et al., 2019b)
Strawberry crinkle virus (SCV)	<i>Alphacytorhabdovirus fragariarugosus</i>	SCRV00	(EFSA Panel on Plant Health et al., 2019b)
Strawberry Kurdistan virus (SKV)	-	not available	(Hajizadeh et al., 2025)
Strawberry latent C virus (SLCV) ³	-	STLCV0	(EFSA Panel on Plant Health et al., 2019b)
Strawberry latent ringspot virus (SLRSV)	<i>Stralarivirus fragariae</i>	SLRSV0	(EFSA Panel on Plant Health et al., 2019b)
Strawberry latent virus (StLV) ⁴	-	not available	(EFSA Panel on Plant Health et al., 2019b)
Strawberry leaf curl virus (StLCV) ³	-	STWLCV	(EFSA Panel on Plant Health et al., 2019b)
Strawberry mild yellow edge virus (SMYEV)	<i>Potexvirus fragariae</i>	SMYEV0	(EFSA Panel on Plant Health et al., 2019b)
Strawberry mottle virus (SMoV)	<i>Sadwavirus fragariae</i>	SMOV00	(EFSA Panel on Plant Health et al., 2019b)
Strawberry necrotic shock virus (SNSV)	<i>Iilarvirus SNSV</i>	SNSV00	(EFSA Panel on Plant Health et al., 2019b)

Name in publication (acronym)	Binomial name	EPPO code	Source
Strawberry pallidosis-associated virus (SPaV)	<i>Crinivirus palidofragariae</i>	SPAV00	(EFSA Panel on Plant Health et al., 2019b)
Strawberry polerovirus 1 (SPV-1)	<i>Polerovirus SPV</i>	SPV100	(EFSA Panel on Plant Health et al., 2019b)
Strawberry pseudo mild yellow edge virus (SPMYEV)	<i>Carlavirus fragariae</i>	SPMYEV	(EFSA Panel on Plant Health et al., 2019b)
Strawberry vein banding virus (SVBV)	<i>Caulimovirus venafragariae</i>	SVBV00	(EFSA Panel on Plant Health et al., 2019b)
Strawberry virus 1 (StrV-1)/Strawberry-associated virus 1 (SaV1)	<i>Alphacytorhabdovirus alphafragariae</i>	not available	(Ding et al., 2019; Fránová et al., 2019)
Strawberry virus 2 (StrV-2)	<i>Alphacytorhabdovirus betafragariae</i>	not available	(Medberry & Tzanetakis, 2022)
Strawberry virus 3 (StrV-3)	<i>Deltanucleorhabdovirus fragariae</i>	not available	(Medberry et al., 2023)
Strawberry virus A (StraVA)	-	not available	(Koloniuk et al., 2022)
Tobacco necrosis virus A (TNVA) ⁵	<i>Alphanecrovirus nicotianae</i>	TNV000	(EFSA Panel on Plant Health et al., 2019b)
Tobacco necrosis virus D (TNVD) ⁵	<i>Betanecrovirus nicotianae</i>	TNV000	(Fránová-Honetšlegrová et al., 1998; Fránová-Honetšlegrová et al., 1999)
Tobacco streak virus (TSV)	<i>Ilarvirus TSV</i>	TSV000	(EFSA Panel on Plant Health et al., 2019b)
Tomato black ring virus (TBRV)	<i>Nepovirus nigranuli</i>	TBRV00	(EFSA Panel on Plant Health et al., 2019b)
Tomato ringspot virus (ToRSV)	<i>Nepovirus lycopersici</i>	TORSV0	(EFSA Panel on Plant Health et al., 2019b)
Turnip yellows virus (TuYV) ⁶	<i>Polerovirus TUYV</i>	TUYV00	(He et al., 2022)

¹ Questionable finding, this virus may have been sequenced from an insect

² Davalos et al. (2021) reported detection of RBDV by dot-immunobinding assay in strawberry crops in the United States. Presence of the virus was not confirmed by other methods. RBDV is a common virus of *Rubus* worldwide and is spread by pollen (Martin, 2001). Therefore, it cannot not be ruled out that the positive detection results were caused by the presence of infected *Rubus*-pollen on the strawberry plants and the status of *Fragaria* as a natural host of RBDV still needs confirmation.

³ SLCV and StLCV are currently considered to be phantom agents (Tzanetakis et al., 2024).

⁴ Virus poorly characterized (EFSA Panel on Plant Health et al., 2019b).

⁵ TNVA and TNVD have been elevated from the rank of strains of tobacco necrosis virus (TNV) to the rank of species and placed in two separate genera (Pringle, 1998). Both species were included as TNV in the pest list compiled by the EFSA Panel on Plant Health et al. (2019b). Older publications that refer to the presence of TNV in strawberry do not always distinguish between the two strains, creating uncertainty about which of the current species is involved. TNVD has since been isolated from asymptomatic plants of *Fragaria x ananassa* (Fránová-Honetšlegrová et al., 1998; Fránová-Honetšlegrová et al., 1999), while there is no definitive prove of the host plant status of *Fragaria* for TNVA.

⁶ Reported as Brassica yellows virus (BrYV) in *Fragaria* by He et al., 2022). However, BrYV is considered an isolate of turnip yellows virus (TuYV, *Polerovirus TUUV*) according to ICTV (2025a).

Annex B Seed transmission of viruses and viroids in *Fragaria* and other hosts

Virus (acronym)	Conclusion – Evidence ¹
Apple mosaic virus (ApMV)	<p>No evidence of seed transmission in <i>Fragaria</i>. Evidence of seed transmission in one other host plant</p> <p>ApMV is listed as seed-transmitted by Tzanetakis & Martin (2013) in a review of strawberry viruses, but without specification of evidence. No other information was found on seed transmission in <i>Fragaria</i>.</p> <p>Results from Cameron & Thompson (1985) and Postman & Mehlenbacher (1992) indicate seed transmission, but no pollen transmission, in hazelnut (<i>Corylus avellana</i>). An experimental study in roses showed no evidence of seed transmission (Golino et al., 2005).</p>
Apple rootstock virus A (ApRVA)	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p> <p>No information was found on seed transmission. ApRVA belongs to the family Rhabdoviridae and members of this family are generally not seed-transmitted (Walker et al., 2026).</p>
Arabis mosaic virus (ArMV)	<p>Evidence of seed transmission in <i>Fragaria</i></p> <p>Seed transmission was found through seeds collected from naturally infected plants of two cultivars of <i>Fragaria x ananassa</i>, from which 11 out of 110 and 2 out of 78 seedlings became infected (Lister and Murant, 1967).</p>
Beet pseudoyellows virus (BPYV)	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p> <p>No information found on seed transmission. Seed transmission is also not expected. BPYB belongs to the family Closteroviridae in which seed transmission is not known (Fuchs et al., 2020).</p>
Beet ringspot virus (BRSV)	<p>Evidence of seed transmission in <i>Fragaria</i>.</p> <p>Seed transmission is reported in <i>Fragaria</i> (Lister & Murant, 1967). The isolate of this study is described as the Scottish isolate of TBRV, isolated from <i>Arctium lappa</i>. This isolate was later sequenced and confirmed to be an isolate of BRSV (Fowkes et al., 2022). Seed transmission of this isolate was found through seeds collected from naturally infected plants of two cultivars of <i>Fragaria x ananassa</i>, from which 26 out of 76 and 2 out of 28 seedlings became infected (Lister & Murant, 1967).</p>
Crassocephalum yellow vein virus (CraYVV)	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p> <p>No information was found on seed transmission. CraYVV belongs to the family of Geminiviridae that are generally phloem-limited (Möller & Maruthi, 2025) and phloem-limited viruses are generally not seed-transmitted (Mink, 1993).</p>
Cucumber mosaic virus (CMV)	<p>No evidence of seed transmission in <i>Fragaria</i>. Evidence of seed transmission in several other host plants.</p> <p>No information was found on seed transmission in <i>Fragaria</i>.</p>

Virus (acronym)	Conclusion – Evidence ¹
	Seed transmission of CMV was (for example) reported in seeds collected from artificially infected <i>Capsicum annuum</i> (Ali & Kobayashi, 2010) and <i>Vigna unguiculata</i> plants (Abdullahi et al., 2001). Seed and pollen transmission was also found in <i>Spinacia oleracea</i> after crossing naturally infected female plants with healthy male plants and vice versa (Yang et al., 1997).
Fragaria chiloensis cryptic virus (FCICV)	<p>Limited or inconclusive evidence of seed transmission in <i>Fragaria</i></p> <p>Seed transmission was deduced from the observation that multiple infected <i>Fragaria chiloensis</i> plants were clones of seedlings grown from seed, combined with the notion that all cryptic viruses are solely seed and pollen transmitted (Tzanetakis et al., 2008).</p> <p>No information was found on seed transmission in other host species (No other host species are reported).</p>
Fragaria chiloensis latent virus (FCILV)	<p>Evidence of seed transmission in <i>Fragaria</i></p> <p>Seed transmission was found through seeds collected from naturally infected <i>Fragaria chiloensis</i> plants (Spiegel et al., 1993), from which 23 out of 42 seedlings became infected.</p>
Fragaria vesca-associated virus 1 (FVaV-1) ²	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p> <p>No information was found on seed transmission. The genomic organization and sequence of this virus indicated relatedness to members of the proposed insect-specific genus Negevirus (Lenz et al. 2020). Evidence that <i>Fragaria</i> is a true natural host plant of this virus is currently lacking.</p>
Hop stunt viroid (HSVd)	<p>No evidence of seed transmission in <i>Fragaria</i>. Evidence of seed transmission in other host plants</p> <p>No information was found on seed transmission in <i>Fragaria</i>.</p> <p>Seed transmission was found in <i>Vitis vinifera</i> and <i>Prunus domestica</i> ('plum' in the publication) (Wan Chow Wah & Symons, 1999; Luigi et al., 2010).</p>
Lycopersicon esculentum nepovirus (LENV)	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p> <p>No information was found on seed transmission. However, many other nepoviruses are known to be seed-transmitted (see also Chapter 4 of the main text).</p>
Olive latent virus 1	<p>No evidence of seed transmission in <i>Fragaria</i>. Evidence of seed transmission in one other host plant</p> <p>Seed transmission has been reported in olive (Saponari et al., 2002).</p>
Plum bark necrosis stem pitting-associated virus (PBNSPaV)	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p> <p>No information was found on seed transmission. Seed transmission is also not expected. PBNSPaV belongs to the family Closteroviridae in which seed transmission is not known (Fuchs et al., 2020).</p>

Virus (acronym)	Conclusion – Evidence ¹
Raspberry bushy dwarf virus (RBDV) ²	<p>Evidence of seed transmission in artificially infected <i>Fragaria</i> plants but evidence on natural host plant status is inconclusive. Evidence of seed transmission in several other host plants</p> <p>Seed transmission was found through seeds collected from artificially infected <i>Fragaria vesca</i> plants (Murant et al., 1974; Credi et al., 1986). Two infected seedlings were found among 64 tested plants by Murant et al. (1974) and eight out of 99 plants by Credi et al. (1986). These experiments demonstrate that, following artificial infection, the virus is able to cause a systemic infection in <i>Fragaria</i>, and infect or contaminate the seeds, which subsequently can result in infected seedlings. However, the status of <i>Fragaria</i> as a natural host needs confirmation (see Viruses and viroids in Annex A). Seed transmission has been demonstrated in plant species known as natural hosts, including <i>Rubus idaeus</i> (Converse, 1973; Murant et al., 1974) and <i>Rubus longanobaccus</i> (Ormerod, 1970).</p>
Raspberry ringspot virus (RpRSV)	<p>Evidence of seed transmission in <i>Fragaria</i></p> <p>Seed transmission was found through seeds collected from naturally infected plants of two cultivars of <i>Fragaria x ananassa</i>, from which 27 out of 55 and 2 out of 24 seedlings became infected (Lister and Murant, 1967).</p>
Spinach latent virus (SpLV)	<p>No evidence of seed transmission in <i>Fragaria</i>. Evidence of seed transmission in several other host plants</p> <p>No information was found on seed transmission in <i>Fragaria</i>.</p> <p>Seed transmission was found in several plant species, including <i>Chenopodium quinoa</i>, <i>Nicotiana clevelandii</i> (Štefanac & Wrischer, 1983), and <i>Spinacia oleracea</i> (Bos et al., 1979;1980).</p>
Strawberry chlorotic fleck-associated virus (SCFaV)	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p> <p>No information was found on seed transmission. Seed transmission is also not expected. SCFaV belongs to the family Closteroviridae in which seed transmission is not known (Fuchs et al., 2020).</p>
Strawberry crinivirus 3 (SCrV-3)	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p> <p>No information was found on seed transmission. Criniviruses are not known to be seed-transmitted (ICTV, 2026a).</p>
Strawberry crinivirus 4 (SCrV-4)	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p> <p>No information was found on seed transmission. Criniviruses are not known to be seed-transmitted (ICTV, 2026a).</p>
Strawberry crinkle virus (SCV)	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p> <p>An experimental study in <i>Fragaria vesca</i> and <i>F. grandiflora</i> showed no evidence of seed transmission (Kacharmazov & Khristov, 1976).</p>

Virus (acronym)	Conclusion – Evidence ¹
	No information was found on seed transmission in other host plants. SCV belongs to the Rhabdoviridae and members of this family are generally not seed-transmitted (Walker et al., 2026).
Strawberry Kurdistan virus (SKV)	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p> <p>No information was found on seed transmission. Seed transmission is also not expected. SKV belongs to the family Closteroviridae in which seed transmission is not known (Fuchs et al., 2020).</p>
Strawberry latent C virus (SLCV) Phantom agent ³	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p> <p>No information was found on seed transmission. Rhabdoviridae are generally not seed-transmitted.</p>
Strawberry latent ringspot virus (SLRSV)	<p>No evidence of seed transmission in <i>Fragaria</i>. Evidence of seed transmission in several other host plants</p> <p>SLRSV is listed as seed-transmitted by Tzanetakis & Martin (2013) in a review of strawberry viruses, but without specification of evidence. No other information was found on seed transmission in <i>Fragaria</i>.</p> <p>Seed transmission was found in several other plant species, including <i>Chenopodium quinoa</i> (Allen et al., 1970) and <i>Petroselinum crispum</i> (Bellardi & Bertaccini, 1992).</p>
Strawberry latent virus (StLV)	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p> <p>No information was found on seed transmission. The virus is poorly characterized (EFSA Panel on Plant Health et al., 2019b)</p>
Strawberry leaf curl virus (StLCV) Phantom agent ³	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p> <p>No information was found on seed transmission.</p>
Strawberry mild yellow edge virus (SMYEV)	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p> <p>No information was found on seed transmission.</p>
Strawberry mottle virus (SMoV)	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p> <p>An experimental study in <i>Fragaria vesca</i> and <i>F. grandiflora</i> showed no evidence of seed transmission (Kacharmazov and Khristov, 1976).</p> <p>No information was found on seed transmission in other host plants.</p>
Strawberry necrotic shock virus (SNSV)	<p>Evidence of seed transmission in <i>Fragaria</i></p> <p>Seed and pollen transmission was found after crossing naturally infected female plants with healthy male plants and vice versa (Johnson et al., 1984). When six crosses of infected female plants and healthy male plants were tested, on average 24% of the seedlings became infected (57 out of 233 seedlings). The disease in the paper by Johnson et al. (1984) is described as</p>

Virus (acronym)	Conclusion – Evidence ¹
	'necrotic shock disease' caused by tobacco streak virus (TSV). At that time, SNSV was considered a strain of TSV, but this virus is now considered a distinct species and the causative agent of necrotic shock disease (Tzanetakis et al., 2004b).
Strawberry pallidosis-associated virus (SPaV)	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p> <p>An experimental study in <i>Fragaria</i> showed no evidence of seed transmission (Tzanetakis et al., 2004a).</p> <p>No information was found on seed transmission in other host plants. Seed transmission is also not expected. SPaV belongs to the family Closteroviridae in which seed transmission is not known (Fuchs et al., 2020).</p>
Strawberry polerovirus 1 (SPV-1)	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p> <p>No information was found on seed transmission. Seed transmission is also not expected. SPV-1 belongs to the genus polerovirus and members of this genus are phloem-limited (Delfosse et al., 2021). Phloem-limited viruses are generally not seed-transmitted (Mink, 1993).</p>
Strawberry pseudo mild yellow edge virus (SPMYEV)	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p> <p>No information was found on seed transmission.</p>
Strawberry vein banding virus (SVBV)	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p> <p>No information was found on seed transmission. Seed transmission of caulimoviruses has not been recorded (ICTV, 2026b).</p>
Strawberry virus 1 (StrV-1)/Strawberry-associated virus 1 (SaV1)	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p> <p>No information was found on seed transmission. StrV-1/SaV1 belongs to the Rhabdoviridae and members of this family are generally not seed-transmitted (Walker et al., 2026).</p>
Strawberry virus 2 (StrV-2)	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p> <p>No information was found on seed transmission. StrV-2 belongs to the Rhabdoviridae and members of this family are generally not seed-transmitted (Walker et al., 2026).</p>
Strawberry virus 3 (StrV-3)	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p> <p>No information was found on seed transmission. StrV-3 belongs to the Rhabdoviridae and members of this family are generally not seed-transmitted (Walker et al., 2026).</p>
Strawberry virus A (StraVA)	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p> <p>No information was found on seed transmission.</p>
Tobacco necrosis virus A (TNVA) ²	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p>

Virus (acronym)	Conclusion – Evidence ¹
	No information was found on seed transmission. No conclusive evidence that <i>Fragaria</i> is a natural host plant (see Annex A).
Tobacco necrosis virus D (TNVD)	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p> <p>No information was found on seed transmission.</p>
Tobacco streak virus (TSV)	<p>No evidence of seed transmission in <i>Fragaria</i>. Evidence of seed transmission in several other host plants</p> <p>Seed transmission of TSV is reported in <i>Fragaria</i> (Johnson et al., 1984), but all references to TSV in <i>Fragaria</i> may actually refer to SNSV (Tzanetakis, 2012), which was initially considered a strain of TSV, but was described in 2004 as a separate species (Tzanetakis et al., 2004b).</p> <p>Seed transmission is reported in several other hosts, e.g. <i>Glycine max</i> (Golnaraghi et al., 2004), <i>Phaseolus vulgaris</i>, <i>Vigna angularis</i> (Kaiser et al., 1991), and <i>Verbesina encelioides</i> (Sharman et al., 2015).</p>
Tomato black ring virus (TBRV)	<p>No evidence of seed transmission in <i>Fragaria</i>. Evidence of seed transmission in several other host plants</p> <p>Seed transmission of TBRV is reported in <i>Fragaria</i> (Lister & Murrant, 1967), but the isolate of this study is described as the Scottish isolate of TBRV, isolated from <i>Arctium lappa</i>. This isolate was later sequenced and confirmed to be an isolate of BRSV (Fowkes et al., 2022). No other information was found in seed transmission in <i>Fragaria</i>.</p> <p>Seed transmission was found in several other hosts, e.g. <i>Beta vulgaris</i> (Gibbs & Harrison, 1964; Murrant, 1983), <i>Rubus idaeus</i> (Lister & Murrant, 1967) and <i>Sambucus nigra</i> (Schimanski, 1987).</p>
Tomato ringspot virus (ToRSV)	<p>Limited or inconclusive evidence of seed transmission in <i>Fragaria</i>. Evidence of seed transmission in several other host plants</p> <p>Seed transmission of ToRSV in <i>Fragaria</i> is mentioned in several publications (Converse & Stace-Smith, 1987; Converse, 1990; Tzanetakis & Martin, 2013). However, the only experimental study found is from Mellor & Stace-Smith (1963). They found seed transmission of a 'ringspot virus' in <i>Fragaria vesca</i> and <i>F. x ananassa</i> but the identity of the virus was uncertain. High transmission rates were observed, with 175 out of 300 seedlings being infected. Conflicting results were obtained regarding the identification of the virus, which may have either been ToRSV or tobacco ringspot virus (TRSV). Therefore, the evidence is considered inconclusive although the virus seems most likely ToRSV and not TRSV. <i>Fragaria vesca</i> is mentioned as a non-host in TRSV-transmission studies of (McGuire, 1973) and <i>Fragaria</i> is not listed as a host of TRSV by both EPPO (2025a) and CABI (2024), which suggest that ToRSV was the virus in the study of Mellor & Stace-Smith (1963).</p> <p>Seed transmission has been found in several other hosts, e.g. <i>Glycine max</i> (Golnaraghi et al., 2004), <i>Rubus idaeus</i> (Braun &</p>

Virus (acronym)	Conclusion – Evidence ¹
	Keplinger, 1973), and <i>Taraxacum officinale</i> (Mountain et al., 1983).
Turnip yellows virus (TuYV)	<p>No evidence of seed transmission in <i>Fragaria</i> and other host plants</p> <p>No information was found on seed transmission. Belongs to the genus polerovirus and members of this genus are phloem-limited (Delfosse et al., 2021). Phloem-limited viruses are generally not seed-transmitted (Mink, 1993).</p>

¹ Evidence for seed transmission in host plants other than *Fragaria* was only investigated if no evidence for seed transmission in *Fragaria* was found.

² Uncertain whether *Fragaria* is a natural host of the virus (see Annex A)

³ Pathogenic agent named in literature but without evidence of its existence (Tzanetakis et al., 2024)